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**PRODUCTION OF 2'-DEOXYNUCLEOSIDES AND 2'-DEOXYNUCLEOSIDE
PRECURSORS FROM 2-DEHYDRO-3-DEOXY-D-GLUCONATE**

This invention relates to a process for preparing 2'-deoxynucleoside compounds or 2'-deoxynucleoside precursors using 2-dehydro-3-deoxy-D-gluconic acid (usually abbreviated as KDG) or its salts as a starting material. A variety of 2'-deoxynucleosides and their analogues are used as a starting material for synthesis or drug formulation in production of an antiviral, anticancer or antisense agent.

Specifically, the invention relates to a method in which KDG or a derivative of KDG is subjected to a decarboxylation step to remove the original carboxy group of KDG. In a preferred embodiment, the KDG used in the method according to the invention is enzymatically produced from D-gluconate or D-glucosamine.

2'-deoxynucleosides and 2'-deoxynucleoside precursors including 2-deoxy-D-ribose are used as starting material for synthesis or drug formulation, for instance, in production of antiviral and anticancer agent. 2'-deoxynucleosides or derivatives thereof and 2'-deoxynucleoside precursors are also used as reagents for research, diagnosis and synthesis of therapeutic antisense molecules.

In one method of the prior art, deoxynucleosides are generated from biological materials such as testis (WO 99/49074) or yeast or fish sperm by enzymatic cleavage of DNA. This method, however, involves several disadvantages, in particular regarding difficulties of obtaining the starting material in sufficient quantity and quality.

The main production process of 2-deoxy-D-ribose currently consists in chemical hydrolysis of DNA. In this case, the deoxyribosyl moiety originates in ribonucleotide reductase activity. No synthesis of 2-deoxy-D-ribose from KDG has been yet described.

In most living cells, deoxyribonucleosides result from a "salvage pathway" of the

nucleotide metabolism. The deoxyribose moiety of deoxyribonucleosides is obtained through the reduction of a ribosyl moiety into di- or triphosphate ribonucleotides catalyzed by ribonucleotide reductases. However, the deoxyribose moiety is not recycled, but is degraded into D-glyceraldehyde-3-phosphate and acetaldehyde following the reactions of central metabolism:

- deoxynucleoside is cleaved into deoxyribose-1-phosphate and nucleobase through phosphorolysis mediated by products of the genes encoding thymidine phosphorylase (deoA), purine-nucleoside phosphorylase (deoD), uridine phosphorylase (udp) or xanthosine phosphorylase (xapA).
- deoxyribose-1-phosphate is converted into deoxyribose-5-phosphate through a reaction catalyzed by deoxyribose phosphate mutase (deoB),
- which is further degraded to D-glyceraldehyde-3-phosphate and acetaldehyde through a reaction catalyzed by deoxyribose-5-phosphate aldolase (deoC).

It has been shown that the deo enzymes also catalyze in vitro the reverse anabolic reactions: Deoxyribose-5-phosphate is obtained in vitro in the presence of purified *Escherichia coli* or *Lactobacillus plantarum* deoxyribose aldolase starting from acetaldehyde and D-glyceraldehyde-3-phosphate (Rosen et al., J. Biol. Chem., 240, (1964), 1517-1524; Pricer, J. Biol. Chem., 235, (1960), 1292-1298). Deoxyribose can also be obtained with acetaldehyde and glyceraldehyde as enzyme substrates, but only with a very low yield (Barbas, J. Am. Chem. Soc. 112 (1990), 2013-2014).

The patent application WO 01/14566 describes the enzymatic synthesis of deoxynucleosides starting from deoxyribose-1-phosphate through the combined activities of three enzymes of the deo operon, i.e. deoxyribose aldolase, deoxyribomutase and phosphorylase (thymidine or purine nucleoside phosphorylase) in a one-pot reaction, using as starting substrates glyceraldehyde-3-phosphate, acetaldehyde and a nucleobase. D-glyceraldehyde-3-phosphate can be obtained from fructose-1,6-bisphosphate by an enzymatic process.

The patent application EP 1179598 describes the use of phosphorylase to catalyze the enzymatic production of deoxynucleosides starting from deoxyribose-1-phosphate and nucleobase. The yield of deoxynucleoside synthesis is improved by

precipitation of phosphate.

However, methods using enzymes of the deo operon working in the reverse direction compared to their biological function show low yields, which indicates serious drawbacks for their use.

In view of the above-described ineffectiveness of the currently applied processes for producing deoxynucleosides and deoxynucleoside precursors, it is an object of the present invention to provide means and methods for the biosynthetic production of deoxynucleosides and deoxynucleoside precursors starting from cheap and commercially available compounds without being dependent on unreliable natural sources.

In particular, there is a need for alternative methods for the production of deoxynucleosides and deoxynucleoside precursors which allow efficient and economical synthesis of deoxyribonucleosides, by means of which the drawbacks of prior art processes are eliminated.

The present invention relates to a method for producing 2'-deoxynucleosides and precursors thereof starting from 2-dehydro-3-deoxy-D-gluconic acid (KDG) or its salts and comprising a decarboxylation step.

In particular, this method is useful for producing 2-deoxy-D-ribose (DRI) as well as synthetically versatile enamine derivatives of DRI as 2'-deoxynucleoside precursors.

The decarboxylation step takes place by reacting either KDG or its salts directly, or a derivative of KDG, usually to cleave the C1-C2 bond of the KDG.

In one embodiment of the invention, KDG or one of its salts undergoes (oxidative) decarboxylation leading to 2-deoxy-D-ribonic acid (DRN) or its salts, itself being further converted into 2-deoxy-D-ribose (DRI) or 2-deoxy-D-ribitol (DRL).

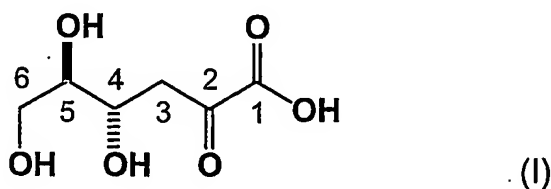
In another embodiment of the invention, decarboxylation takes place by reacting KDG or its salts with an amine, leading to an enamine derivative. This high energy enamine derivative can be further converted into DRI by hydrolysis.

In another embodiment of the invention, (oxidative) decarboxylation is carried out on 3-deoxy-D-gluconic acid (DGN) or its salts and/or 3-deoxy-D-mannonic acid (DMN) or its salts as derivatives of KDG, leading to DRI. Production of a mixture of DGN and DMN takes place by reduction of KDG. The decarboxylation is preferably carried out via reaction with hydrogen peroxide.

In another embodiment of the invention, (oxidative) decarboxylation is carried out on 3-deoxy-D-glucosaminic acid (DGM) or its salts and/or 3-deoxy-D-mannosaminic acid (DMM) or its salts, leading to DRI. Production of a mixture of DGM and DMM takes place from KDG by reductive amination.

Another aspect of the invention is a convenient and cost-effective method for preparing KDG or its salts to be used in the above methods. This method starts either from D-gluconate or from D-glucosamine through the use of recombinant enzymes. The invention provides a novel nucleotide sequence encoding a polypeptide having D-gluconate dehydratase activity and a nucleotide sequence encoding a polypeptide having D-glucosamine deaminase activity.

The starting material used for the method of the present invention is KDG, represented by formula (I) below or one of its salts, or a protected derivative thereof wherein one or more of the hydroxyl groups at positions 4, 5 and/or 6 are protected by a protection group known in the art.



The term "2'-deoxynucleoside" as used herein relates to 2'-deoxyribonucleosides which are N-glycosides, and wherein the basic N-atom of the nucleobase or nucleobase analog is bound to the anomeric carbon atom of 2-deoxy-D-ribose, or one of its derivatives. Examples of a suitable nucleobase are adenine, cytosine, guanine, thymine, uracil, 2,6-diaminopurine, and hypoxanthine. Examples of nucleobase analogs are 5-azacytosine, 2-chloro-adenine, 5-iodo-cytosine, 8-azaguanine, 5-iodo-uracil, 5-bromo-uracil, 5-fluoro-uracil, 5-ethyl-uracil and 5-trifluoromethyl-uracil.

The term "2'-deoxynucleoside precursors" as used herein, relates to compounds which can be easily converted into 2'-deoxynucleosides by applying methods known in the prior art. Preferred 2'-deoxynucleoside precursors are 2-deoxy-D-ribose (DRI) or carbohydrate compounds which can be converted into the 2-deoxy-D-ribosyl moiety of 2'-deoxynucleosides, for instance, those established in the prior art 1-phospho-2-deoxy-D-ribose, 5-phospho-2-deoxy-D-ribose and those established by the present invention 2-deoxy-D-ribitol, 2-deoxy-D-ribonic acid, 2-deoxy-D-ribono-1,4-lactone, 1-N-morpholino-3,4,5-trihydroxy-pentene-1, and their derivatives.

The method of the invention encompasses methods wherein the decarboxylation step is directly carried out on KDG or its salts or on compounds derived from KDG. Preferred KDG derivatives are 3-deoxy-D-gluconic acid, 3-deoxy-D-mannonic acid, 3-deoxy-D-glucosaminic acid and 3-deoxy-D-mannosaminic acid and their respective salts.

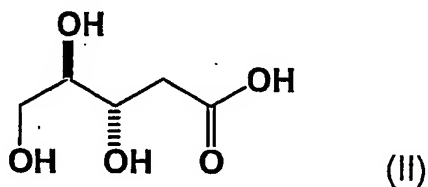
Furthermore, KDG and its salts or protected forms of these wherein one or more of the hydroxyl groups at the positions 4,5 and /or 6 are replaced by protecting groups known for that purpose in the art are also suitable starting materials for the decarboxylation reaction of the present invention. Unless noted otherwise, any reference to KDG in the following specification embraces protected forms of KDG, just as reference to KDG derivatives is intended to embrace protected forms of these derivatives. Similarly, any reference to the products obtained in the methods of the invention is intended to encompass protected forms of these products. Preferred

protection groups for the purpose of the invention are those which replace the respective hydroxyl groups by acetate ester, benzoate ester, allyl ether, benzyl ether, trityl ether, ter-butyldimethylsilyl (TBDMS) ether, isopropylidene or a benzylidene acetal.

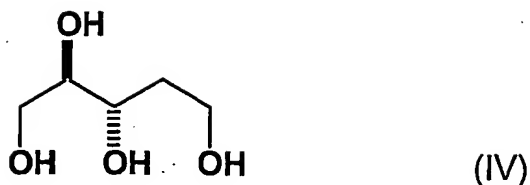
It should be understood that, depending on suitable reaction conditions for the embodiments of the invention, the carboxylic groups contained in the organic acids used as reactants or obtained as products can be in a protonated form or in their salt form, or may be present in equilibrium. Exemplary salts of these acids are those which have metal or ammonium ions as counterions, particularly alkali metal ions such as sodium and/or potassium.

Most of the carbohydrate compounds and their derivatives described in the present invention exist under several cyclic form but for simplicity reasons have been represented by open chain formulas. It is understood that the present invention encompasses all these isomeric or tautomeric forms.

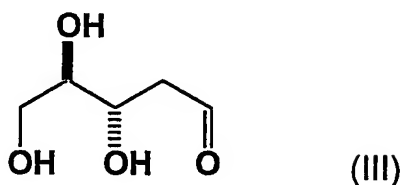
In a first embodiment of the invention, KDG or its salts is reacted with hydrogen peroxide and undergoes (oxidative) decarboxylation to 2-deoxy-D-ribonic acid (DRN), a compound of formula (II) or its salts.



The product may be further converted into or 2-deoxy-D-ribitol (DRL), represented by formula (IV)



or 2-deoxy-D-ribose (DRI), represented by formula (III)



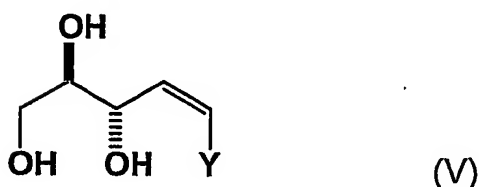
DRN, DRL and particularly DRI are among preferred 2'-deoxynucleoside precursors for the purpose of the present invention. Conversion of DRN to DRI may proceed directly or via DRL as an intermediate.

Preferably, the preparation of DRN is carried out by oxidative decarboxylation of sodium or potassium 2-dehydro-3-deoxy-D-gluconate in aqueous solution with hydrogen peroxide at room temperature as described in example 5. A general method for the preparation of aldonic acids by oxidative decarboxylation of 2-ketoaldonic acids is described in patent EP 1 038 860 A1.

Preferably, the preparation of DRL is carried out by hydrogenation of 2-deoxy-D-ribonolactone in aqueous solution with Rhodium catalyst on carbon at a temperature of 130°C under a pressure of 80 bars as described in example 6. 2-Deoxy-D-ribonolactone can be easily prepared by converting a 2-deoxy-D-ribonate (DRN salt) into 2-deoxy-D-ribonic acid, which is in equilibrium with its lactonic form in aqueous solutions (Han, Tetrahedron. 1993. 49, 349-362; Han, Tetrahedron Asymmetry. 1994. 5, 2535-62).

Preferably the preparation of 2-deoxy-D-ribose (DRI) is carried out by oxidization of 2-deoxy-D-ribitol (DLR), e.g. with chromium oxide in pyridine.

In another embodiment of the invention, decarboxylation takes place by reacting (KDG) or its salts with an amino group-containing reagent Y-H leading to a compound of formula (V).



or its respective trans isomer or a protected form thereof, as a 2'-deoxynucleoside precursor. Y-H represents an amine with the hydrogen atom H bound to the nitrogen of the amino group.

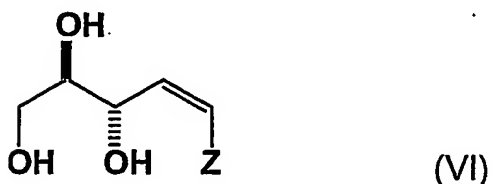
In a preferred embodiment of the invention, the amino group-containing reagent represented by Y-H is a linear or cyclic secondary amine; a primary amine that possess a β -carbonyl group, preferably 3-amino-2-indolinone which was found to be effective for the decarboxylation of α -keto acids (Hanson, J. Chem. Education, 1987, 591-595). In each of these cases, -Y in formula (V) represents the respective nitrogen containing residue derived from these amino-group containing reagent.

Preferably, the compound of formula (V) represents an enamine produced via reaction of a linear or cyclic secondary amine as Y-H.

Preferred cyclic secondary amines are morpholine, pyrrolidine, piperidine, or N-methyl piperazine; preferred non-cyclic amines are those of the formula $R_1\text{-NH-R}_2$, wherein R_1 and R_2 independently represent a linear or branched alkyl group of 1-8, preferably 1 to 4 carbon atoms. Particularly preferred as a non-cyclic amine is diethylamine.

Particularly preferred as a cyclic amine is morpholine.

The compound of formula (V) or its trans isomer or a protected form thereof can be further reacted with Z-H, wherein H represents a hydrogen atom and Z represents a leaving group, to produce a compound of formula (VI)

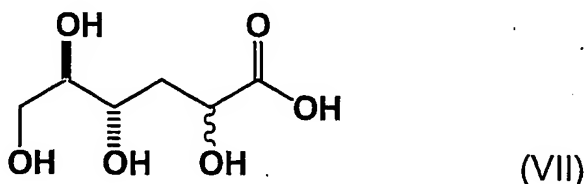


or its respective trans isomer or a protected form thereof, as a 2'-deoxynucleoside precursor. Z-H is preferably water, in which case the compound of formula (VI) is DRI or a protected form thereof (keto-enol-tautomerism).

Preferably, the preparation of the compound of formula (V) is carried out by reacting KDG in benzene with the amine, e.g. morpholine under reflux using the method described in example 7, leading to 1-N-morpholino-3,4,5-trihydroxy-pentene-1. Acid catalysed hydrolysis with water yields 2-deoxy-D-ribose (DRI)

A general route to aldehydes via enamines from α -oxocarboxylic acids carrying β -hydrogens is described by Stamos (Tetrahedron Lett. 23 (1982), 459-462). Other methods for the preparation and hydrolysis of enamines have been described elsewhere (Stork, J. Am. Chem. Soc. 85 (1963), 207-222; Stamhuis, J. Org. Chem. 30 (1965), 2156-2160).

In another embodiment of the invention, KDG or its salt is converted to 3-deoxy-D-gluconic acid (DGN) and/or 3-deoxy-D-mannonic acid (DMN) represented by formula (VII) or the salts of these compounds

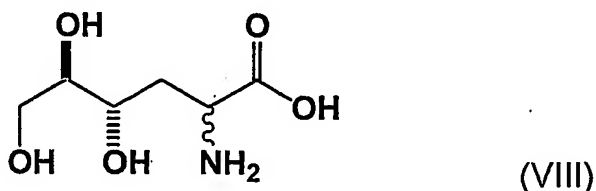


The products resulting from this reaction undergo (oxidative) decarboxylation, preferably using hydrogen peroxide, to yield DRI. Production of a mixture of DGN and DMN or their salts takes place from KDG or its salts by reduction.

Preferably the preparation of 2-deoxy-D-ribose (DRI) is carried out by non-stereoselective reduction of 2-dehydro-3-deoxy-D-gluconic acid in water with sodium borohydride at room temperature using the method described for 2-keto-3-deoxyheptonic acid by Weissbach (J. Biol. Chem. 234 (1959), 705-709), followed by oxidative decarboxylation of 3-deoxy-D-gluconate and 3-deoxy-D-mannonate with hydrogen peroxide as described e.g. in US patent 3,312,683; Richards J. Chem. Soc. (1954), 3638-3640; Sowden J. Am. Chem. Soc. 76 (1954), 3541-3542.

In another preferred embodiment, the preparation of a mixture of DGN and DMN is carried out by hydrogenation of 2-dehydro-3-deoxy-D-gluconate in aqueous solution with 6% mol Nickel Raney catalyst or Platinum oxide at room temperature under a pressure of 6 bars.

In another embodiment of the invention, KDG or its salt is converted to 3-deoxy-D-glucosamine (DGM) or 3-deoxy-D-mannosamine (DMM) represented by formula (VIII) or the salts of these compounds



The products resulting from this reaction undergo (oxidative) decarboxylation, preferably using ninhydrin, to yield DRI. Production of a mixture of DGM and DMM or their salts takes place from KDG or its salts by reductive amination.

Preferably the preparation of 2-deoxy-D-ribose is carried out by non-stereoselective reductive amination of sodium or potassium 2-dehydro-3-deoxy-D-gluconate in

aqueous solution with ammonia and sodium cyanoborohydride at room temperature, followed by oxidative decarboxylation of 3-deoxy-D-2-glucosamine and 3-deoxy-D-2-mannosamine with ninhydrin using the method described for the synthesis of 2-deoxy-D-allose by Shelton (J. Am. Chem. Soc. 118 (1996), 2117-2125; and Borch, J. Am. Chem. Soc. 93 (1971), 2897; Durrwachter, J. Am. Chem. Soc. 108 (1986), 7812 referenced therein).

Furthermore, the present invention provides a method for producing the compound of formula (III) (2-deoxy-D-ribose) by converting the compound of formula (I) or one of its salts (KDG) in one single step. Preferably this conversion is achieved through enzymatic catalysis. This conversion is preferably catalysed by a keto acid decarboxylase. Preferred keto acid decarboxylases are thiamin pyrophosphate (TPP) dependent keto acid decarboxylases. Examples of TPP dependent keto acid decarboxylases are pyruvate decarboxylase (EC 4.1.1.1), a benzoylformate decarboxylase (EC 4.1.1.7), an indolepyruvate decarboxylase (EC 4.1.1.74), a phosphonopyruvate decarboxylase, a sulfopyruvate decarboxylase (EC 4.1.1.79), an oxalyl-coenzymeA decarboxylase (EC 4.1.1.8), an oxoglutarate decarboxylase (EC 4.1.1.71) or a phenylpyruvate decarboxylase (EC 4.1.1.43). It could be shown that keto acid decarboxylases, e.g., pyruvate decarboxylase enzymes from different organisms, can convert KDG into 2-deoxy-D-ribose (see Examples 8 to 12). In principle any keto acid decarboxylase can be used in connection with the present invention.

In a preferred embodiment of the method according to the invention KDG is converted into 2-deoxy-D-ribose by use of an enzyme having pyruvate decarboxylase activity.

A pyruvate decarboxylase catalyses the following reaction:



Several pyruvate decarboxylases (PDC) have been characterized as well as the corresponding *pdc* genes, for instance PDC from *Zymomonas mobilis* (Genbank accession number AAD19711; Neale et al., J. Bacteriol. 1987, 169:1024-1028), PDC from *Saccharomyces cerevisiae* (Genbank accession number NP013145; Candy et al., J. Gen. Microbiol. 1991, 137:2811-2815), PDC from *Acetobacter pasteurianus* (Genbank accession number AAM21208; Raj et al., Arch. Microbiol. 2001, 176:443-451), PDC from *Zymobacter palmae* (Genbank accession number AAM49566; Raj et al., Appl. Environ. Microbiol. 2002, 68:2869-2876), PDC from *Sarcina ventriculi* (Genbank accession number AAL18557; Lowe et al., J. Gen. Microbiol. 1992, 138:803-807). Many other pyruvate decarboxylases seems to occur in plants, fungi and bacteria as evidenced by the occurrence in these organisms of genes sharing sequence homologies with well-established *pdc* genes. Examples of such putative pyruvate decarboxylases are:

PDC from plants:

Arabidopsis thaliana (Genbank accession number T48155)
Echinochloa crus-galli (Genbank accession number AAM18119)
Oryza sativa (Genbank accession number NP922014)
Rhizopus oryzae (Genbank accession number AAM73540)
Lotus corniculatus (Genbank accession number AAO72533)
Zea mays (Genbank accession number BAA03354)
Pisum sativum (Genbank accession number CAA91445)
Garden pea (Genbank accession number S65470)
Nicotiana tabaccum (Genbank accession number CAA57447)
Solanum tuberosum (Genbank accession number BAC23043)
Fragaria ananassa (Genbank accession number AAL37492)
Cucumis melo (Genbank accession number AAL33553)
Vitis vinifera (Genbank accession number AAG22488)

PDC from Fungi :

Saccharum officinarum (Genbank accession number CAB61763)
Aspergillus oryzae (Genbank accession number AAD16178)
Aspergillus parasiticus (Genbank accession number P51844)
Saccharomyces cerevisiae (Genbank accession number NP013145)
Flammulina velutipes (Genbank accession number AAR00231)

Saccharomyces kluyveri (Genbank accession number AAP75899)
Schizosaccharomyces pombe (Genbank accession number CAB75873)
Candida glabrata (Genbank accession number AAN77243)
Neurospora crassa (Genbank accession number JN0782)
Pichia stipitis (Genbank accession number AAC03164)
Kuyveromyces lactis (Genbank accession number CAA61155)
Emericella nidulans (Genbank accession number AAB63012)

PDC from Prokaryotes:

Mycobacterium bovis (Genbank accession number CAD93738)
Mycobacterium leprae (Genbank accession number CAC31122)
Mycobacterium tuberculosis (Genbank accession number NP215368)
Mycoplasma penetrans (Genbank accession number NP758077)
Clostridium acetobutylicum (Genbank accession number NP149189)
Acetobacter pasteurianus (Genbank accession number AAM21208)
Zymobacter palmae (Genbank accession number AAM49566)
Zymomonas mobilis (Genbank accession number AAD19711)
Sarcina ventriculi (Genbank accession number AAL18557)
Nostoc punctiforme (Genbank accession number ZP00110850)

Such enzymes can be easily produced by recombinant microorganisms overexpressing the corresponding gene. Examples of genes coding for TPP dependent keto acid decarboxylases are pdc from *Zymomonas mobilis* (Genbank accession number AF124349), pdc from *Saccharomyces cerevisiae* (Genbank accession number NC001144), pdc from *Acetobacter pasteurianus* (Genbank accession number AF368435), pdc from *Zymobacter palmae* (Genbank accession number AF474145), pdc from *Sarcina ventriculi* (Genbank accession number AF354297). Other pdc genes can be found at Genbank corresponding to the above list of putative pyruvate decarboxylases.

In a preferred embodiment the pyruvate decarboxylase is of eukaryotic origin, more preferably it is from yeast and most preferably it is from *Saccharomyces cerevisiae*. In a particularly preferred embodiment the pyruvate decarboxylase is the pyruvate decarboxylase from *S. cerevisiae* which has the amino acid sequence as shown in SEQ ID NO: 21 (see also GenBank accession number NP013145).

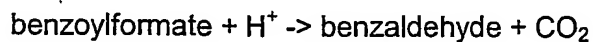
In another preferred embodiment the pyruvate decarboxylase is of prokaryotic origin, more preferably it is from an organism of the genus *Zymomonas* and most preferably from *Zymomonas mobilis*. In a particularly preferred embodiment the pyruvate decarboxylase is the pyruvate decarboxylase from *Z. mobilis* which has the amino acid sequence as shown in SEQ ID NO: 19 (see also GenBank accession number AAD19711).

In another preferred embodiment the prokaryotic pyruvate decarboxylase is from an organism of the genus *Acetobacter*, more preferably from the species *Acetobacter pasteurianus*. Particularly preferred the pyruvate decarboxylase is that of *A. pasteurianus* which shows the amino acid sequence as given in SEQ ID NO: 25 (see also GenBank accession number AAM21208).

In a further preferred embodiment the pyruvate decarboxylase is from an organism of the genus *Zymobacter*, more preferably of the species *Zymobacter palmae*. Particularly preferred is a pyruvate decarboxylase from *Z. palmae* which shows the amino acid sequence given in SEQ ID NO: 29 (see also GenBank accession number AAM49566).

In another preferred embodiment of the method according to the invention KDG is converted into 2-deoxy-D-ribose by use of an enzyme having benzoylformate decarboxylase activity.

A benzoylformate decarboxylase catalyses the following reaction:



A benzoylformate decarboxylase (BDC) from *Pseudomonas putida* (Genbank accession number AAC15502; Tsou et al., *Biochemistry*. 1990, 29:9856-9862) has been characterized as well as the corresponding gene *mdlC* (Genbank accession number AY143338). This enzyme has been shown to decarboxylate both D and L isomers of 2-keto-4,5-dihydroxyvalerate into the respective isomers of 3,4-

dihydroxybutanal (Niu et al., J. Am. Chem. Soc. 125 (2003), 12998-12999). Many other benzoylformate decarboxylases seems to occur in bacteria and archaeobacteria as evidenced by the occurrence in these organisms of genes sharing sequence homologies with genes coding for well-established BDC. Examples of such putative benzoylformate decarboxylases are:

BDC from bacteria:

Pseudomonas aeruginosa (Genbank accession number NP_253588)

Rhodopseudomonas palustris (Genbank accession number NP_946955)

Streptomyces coelicolor (Genbank accession number NP_631486)

Chromobacterium violaceum (Genbank accession number NP_902771)

Bradyrhizobium japonicum (Genbank accession number NP_774243)

BDC from archaeobacteria:

Sulfolobus solfataricus (Genbank accession number NP_343070)

Thermoplasma acidophilum (Genbank accession number NP_393976)

Thermoplasma volcanium (Genbank accession number NP_111716)

Such enzymes can be easily produced by recombinant microorganisms overexpressing the corresponding bdc gene. Such genes can be found at Genbank corresponding to the above list of putative benzoylformate decarboxylases.

Another example for a thiamine dependent decarboxylase which can be used in the method according to the invention is phosphonopyruvate decarboxylase. Several phosphonopyruvate decarboxylases (PPD) have been characterized as well as the corresponding genes, for instance PPD from *Bacteroides fragilis* (Genbank accession number AAG26466; Zhang et al., J. Biol. Chem. 2003, 278:41302-41308), PPD from *Streptomyces wedmorensis* (Genbank accession number BAA32496; Nakashita et al., J. Antibiot. 1997, 50:212-219). Many other phosphonopyruvate decarboxylases seem to occur in bacteria as evidenced by the occurrence in these organisms of genes sharing sequence homologies with genes coding for well-established PPD. Examples of such putative phosphonopyruvate decarboxylases are: PPD from *Bacteroides thetaiotaomicron* (Genbank accession number NP_810632), PPD from *Amycolatopsis orientalis* (Genbank accession number CAB45023), PPD from *Clostridium tetani* E88 (Genbank accession number

NP_782297), PPD from *Streptomyces viridochromogenes* (Genbank accession number CAA74722), PPD from *Streptomyces hygroscopicus* (Genbank accession number BAA07055), PPD from *Streptomyces coelicolor* A3 (Genbank accession number NP_733715), *Streptomyces rishiriensis* (Genbank accession number AAG29796), *Bordetella pertussis* (Genbank accession number CAE 41214. Such enzymes can be easily produced by recombinant microorganisms overexpressing the corresponding gene.

A further example of a thiamine dependent decarboxylases which can be used in the method according to the present invention is sulfopyruvate decarboxylase. A sulfopyruvate decarboxylases (SPD) from *Methanococcus jannaschii* (Graupner et al., J. Bacteriol. 2000. 182:4862-4867) consisting of two subunits ComD (Genbank accession number P58415) and ComE (Genbank accession number P58416) has been characterized as well as the corresponding genes. Many other sulfopyruvate decarboxylases seems to occur in archaeobacteria and in bacteria as evidenced by the occurrence in these organisms of genes sharing sequence homologies with genes coding for well-established SPD.

Another further example of thiamine dependent decarboxylase which can be used in the method according to the present invention is indolepyruvate decarboxylase. Several indolepyruvate decarboxylases (IPD) have been characterized as well as the corresponding genes, for instance IPD from *Enterobacter cloacae* (Genbank accession number BAA14242; Scutz et al., 2003, Eur. J. Biochem. 270:2322-2331), IPD from *Azospirillum brasilense* (Genbank accession number AAC36886; Costacurta et al., Mol. Gen. Genet. 1994, 243:463-472), IPD from *Erwinia herbicola* (Genbank accession number AAB06571; Brandl et al., Appl. Environ. Microbiol. 1996, 62:4121-4128). Many other indolepyruvate decarboxylases seem to occur in bacteria as evidenced by the occurrence in these organisms of genes sharing sequence homologies with genes coding for well-established IPD.

Still another further example of a thiamine dependent decarboxylases which can be used in the method according to the present invention is phenylpyruvate decarboxylase. A phenylpyruvate decarboxylase from yeast (Genbank accession

number NP010668; Vuralhan et al., Appl. Environ. Microbiol. 2003, 69:4534-41) has been characterized as well as the corresponding gene ARO10 (Genbank accession number NC001136).

In a preferred embodiment of the method according to the invention in which the decarboxylation step is effected by an enzymatic reaction, the pH value is regulated by addition of an acid to be between pH 5 and pH 9, preferably between pH 6 and pH 8. In principle, any suitable acid can be used for this purpose. Preferred acids are HCl, H₂SO₄, D-gluconic acid or 2-dehydro-3-deoxy-D-gluconic acid.

Another aspect of the invention is a convenient and cost-effective method for preparing KDG either from D-gluconate (GCN) or from D-glucosamine through the use of recombinant enzymes.

In a preferred embodiment of the method of the invention, the compound of formula (I) is produced in a preliminary step from a D-gluconate salt by the use of a D-gluconate dehydratase activity. Preferred salts are potassium or sodium D-gluconate. Preferably the D-gluconate dehydratase is encoded by a polynucleotide comprising the nucleotide sequence selected from the group consisting of:

- (a) nucleotide sequences encoding a polypeptide comprising the amino acid sequence of SEQ ID N°2;
- (b) nucleotide sequences comprising the coding sequence of SEQ ID N°1;
- (c) nucleotide sequences encoding a fragment encoded by a nucleotide sequence of (a) or (b);
- (d) nucleotide sequences hybridising with a nucleotide sequence of any one of (a) to (c); and
- (e) nucleotide sequences which deviate from the nucleoside sequence of (d) as a result of degeneracy of the genetic code.

The enzymatic synthesis of KDG or its salts using D-gluconate dehydratase proceeds according to the following reaction: D-gluconate is converted into KDG by the elimination of one water molecule. The activity of a D-gluconate dehydratase has been characterized in different bacterial species e.g. in *Alcaligenes* (Kerstens,

Methods in Enzymology 42 (1975), 301-304); *Clostridium pasteurianum*, (Gottschalk, Methods in Enzymology 90 (1982), 283-287); *Thermoplasma acidophilum* (Budgen, FEBS Letters 196 (1986), 207-210) and *Sulfolobus solfataricus* (Nicolaus, Biotechnology Letters 8(7) (1986), 497-500). The preferred D-gluconate dehydratase was identified by screening several collection strains for D-gluconate dehydratase activity. The gene encoding a D-gluconate dehydratase, which was designated *gcnD* was selected from a genomic library of *Agrobacterium tumefaciens* strain C58, and further inserted in a multi copy vector optimised for expression. It was shown that a crude extract from *E. coli* cells over-expressing the *gcnD* gene catalysed the total conversion of D-gluconate into KDG (see Example 2).

In a further preferred embodiment of the method of the invention, the compound of formula (I) is produced in a preliminary step from D-glucosaminatate by the use of a D-glucosaminatate deaminase activity. Preferably the D-glucosaminatate deaminase is encoded by a polynucleotide comprising the nucleotide sequence selected from the group consisting of:

- (f) nucleotide sequences encoding a polypeptide comprising the amino acid sequence of SEQ ID N°4;
- (g) nucleotide sequences comprising the coding sequence of SEQ ID N°3;
- (h) nucleotide sequences encoding a fragment encoded by a nucleotide sequence of (a) or (b);
- (i) nucleotide sequences hybridising with a nucleotide sequence of any one of (a) to (c); and
- (j) nucleotide sequences which deviate from the nucleoside sequence of (d) as a result of degeneracy of the genetic code.

The enzymatic synthesis of KDG or its salts using D-glucosaminatate deaminase proceeds according to the following reaction: D-glucosaminatate is converted into KDG by the elimination of one molecule water and one molecule of ammonia. The activity of a D-glucosaminatate deaminase has been characterized in different bacterial species e.g. in *Pseudomonas fluorescens* (Iwamoto, Agric. Biol. Chem. 53 (1989), 2563-2569) *Agrobacterium radiobacter* (Iwamoto, FEBS Letters 104 (1979), 131-134;

Iwamoto, J. Biochem. 91 (1982), 283-289), and its requirement for Mn^{2+} ion was shown (Iwamoto, Biosci. Biotech. Biochem. 59 (1995), 408-411).

The preferred D-glucosamine deaminase was identified by screening several collection strains for D-glucosamine deaminase activity. The gene encoding a D-glucosamine deaminase, which was designated gmaA was isolated from *Agrobacterium tumefaciens* strain C58 by cloning a gene annotated as a putative D-serine deaminase. The gmaA gene was further inserted in a multi copy vector optimised for expression. It was shown that a crude extract from *E. coli* cells over-expressing the gmaA gene catalysed the conversion of D-glucosamine into KDG (see Example 4).

In a preferred embodiment the present invention relates to a method for producing a compound of formula III, in particular 2-deoxy-D-ribose, starting from D-gluconate or D-glucosamine by enzymatic reactions which, in a first step, convert D-gluconate or D-glucosamine into KDG as described above and, in a second step, convert KDG into 2-deoxy-D-ribose as described above.

Thus, the enzymatic conversion of D-gluconate into KDG can be achieved by the use of a D-gluconate dehydratase. The enzymatic conversion of D-glucosamine into KDG can be achieved by the use of a D-glucosamine deaminase. With respect to the preferred embodiments the same applies as has already been set forth above.

The enzymatic conversion of the resulting KDG into 2-deoxy-D-ribose can be achieved by the use of a keto acid decarboxylase. With respect to the preferred embodiments the same applies as has been set forth above.

The enzymatic two step method of converting D-gluconate or D-glucosamine into 2-deoxy-D-ribose via KDG can be carried out in vitro by using cell extracts of cells expressing the corresponding enzymes or by using purified or partially purified enzymes. The enzymes can be enzymes which are naturally expressed in an organism or they may be recombinantly produced. Methods of preparing and isolating corresponding (recombinant) enzymes are well-known to the person skilled in the art.

In a preferred embodiment the enzymatic two step method of converting D-gluconate or D-glucosamine into 2-deoxy-D-ribose via KDG is carried out in vivo, i.e. by using a suitable organism, which expresses the required enzyme activities. This organism

may be any type of organism, preferably it is a cell, e.g. a plant, an animal, a fungal cell or a bacterial cell. Most preferably fungal or bacterial cells are used. Preferred fungi are yeasts, such as *Saccharomyces cerevisiae*; preferred bacterial cells are, e.g. *E. coli*, *Zymomonas mobilis*, *Zymobacter palmae*, *Acetobacter pasteurianus*, *Acinetobacter calcoaceticus*, *Agrobacterium tumefaciens* and *Bacillus subtilis*. The organism may be an organism which endogenously already expresses one of the enzymatic activities, i.e. a D-gluconate dehydratase or a D-glucosamine deaminase for producing KDG, or a keto acid decarboxylase for converting KDG into 2-deoxy-D-ribose, and in which the respective other enzymatic activity is expressed due to the introduction of a corresponding exogenous nucleic acid molecule encoding the corresponding enzyme. Alternatively, the organism may also be an organism which naturally does not express the enzyme activities required for converting D-gluconate or D-glucosamine into KDG and further into 2-deoxy-D-ribose and in which corresponding foreign nucleic acid molecules have been introduced encoding D-gluconate dehydratase or D-glucosamine deaminase and a keto acid decarboxylase, respectively.

In a particularly preferred embodiment the organism is an organism which does not express a KDG kinase (*kdgK*) activity. Such an enzyme activity would lead to a phosphorylation of KDG to KDPG, which in turn is cleaved by an aldolase into pyruvate and glyceraldehyde-phosphate, thereby diverting KDG into a different unwanted metabolic pathway. It is possible to use for the method according to the invention organisms which naturally do not express a *kdgK* gene. If the used organism naturally expresses a *kdgK*, means and methods are well-known to the skilled person to produce mutants or variants of such an organism in which the corresponding *kdgK* gene is inactivated.

If the described method according to the invention is carried out in vivo by using an organism which expresses a D-gluconate dehydratase for converting D-gluconate into KDG and a keto acid decarboxylase for converting KDG into 2-deoxy-D-ribose, this has the advantage that one can provide D-gluconate as a substrate in the culture medium used to culture the organism. D-gluconate is taken up by the organism and is converted into 2-deoxy-D-ribose.

In another particularly, preferred embodiment the organism is an organism which does not express a KDG aldolase (encoded by the *eda* gene in *E. coli*) activity. Such an enzyme activity would lead to cleavage of KDG into pyruvate and glyceraldehydes, thereby diverting KDG into a different unwanted metabolic pathway. It is possible to use for the method according to the invention organisms which naturally do not express an *eda* gene. If the used organism expresses an *eda* gene, means and methods are well-known to the skilled person to produce mutants or variants of such an organism in which the corresponding *eda* gene is inactivated.

In still another particularly, preferred embodiment the organism is an organism which does not express a 2-deoxy-D-ribose aldolase (encoded by the *deoC* gene in *E. coli*) activity. Such an enzyme activity would lead to cleavage of 2-deoxy-D-ribose into acetaldehyde and glyceraldehyde, thereby diverting 2-deoxy-D-ribose into a different unwanted metabolic pathway. It is possible to use for the method according to the invention organisms which naturally do not express a *deoC* gene. If the used organism expresses a *deoC* gene, means and methods are well-known to the skilled person to produce mutants or variants of such an organism in which the corresponding *deoC* gene is inactivated. For instance a *deoC* mutant of *E. coli* has been reported (Valentin-Hansen, EMBO J. 1 (1982), 317-322) as well as a method for deleting the *deo* operon in *E. coli* (Kaminski, J. Biol. Chem. 277 (2002), 14400-14407; Valentin-Hansen, Molec. Gen. Genet. 159 (1978), 191-202).

The present invention also relates to organism which are capable of enzymatically converting D-gluconate into KDG due to the expression of a D-gluconate dehydratase and/or of enzymatically converting D-glucosamine into KDG due to the expression of a D-glucosamine deaminase and which are furthermore capable of enzymatically converting KDG into 2-deoxy-D-ribose by a decarboxylation reaction catalysed by a keto acid decarboxylase. The organism may in principle be any suitable organism, preferably, it is a cell, e.g. a plant cell, an animal cell, a fungal cell or a bacterial cell. More preferably, it is a fungal or a bacterial cell. Preferred fungi are yeasts, e.g. *Saccharomyces cerevisiae*. Preferred bacteria are *Escherichia coli*, *Zymomonas mobilis*, *Zymobacter palmae*, *Acetobacter pasteurianus*, *Acinetobacter calcoaceticus*, *Agrobacterium tumefaciens* and *Bacillus subtilis*. In one aspect, the

organism is an organism which already endogenously expresses a D-gluconate dehydratase or a D-glucosamine deaminase and into which a foreign nucleic acid molecule has been introduced which encodes a keto acid decarboxylase which can catalyse the decarboxylation of KDG to 2-deoxy-D-ribose. With respect to the preferred embodiments of the keto acid decarboxylase the same applies as has been set forth previously.

In another aspect, the organism is an organism which already expresses a keto acid decarboxylase which is capable of converting KDG into 2-deoxy-D-ribose by a decarboxylation reaction but which does not naturally express a D-gluconate dehydratase or a D-glucosamine deaminase, and into which a foreign nucleic acid molecule has been introduced which encodes a D-gluconate dehydratase and/or which encodes a D-glucosamine deaminase. I.e. the organism can be genetically modified so as to express a D-gluconate dehydratase or a D-glucosamine deaminase or both enzymes.

In a further aspect, the organism is an organism, which naturally does not express a D-gluconate dehydratase, a D-glucosamine deaminase and a keto acid decarboxylase which is capable of converting KDG by decarboxylation into 2-deoxy-D-ribose, and into which foreign nucleic acid molecules have been introduced encoding a D-gluconate dehydratase or a D-glucosamine deaminase, or both, and a nucleic acid molecule which encodes a keto acid decarboxylase which is capable of converting KDG into 2-deoxy-D-ribose by decarboxylation.

With respect to the preferred embodiments of the D-gluconate dehydratase, the D-glucosamine deaminase and the keto acid decarboxylase to be expressed in the organisms according to the invention, the same applies which has been set forth above in connection with the method according to the invention.

In a particularly preferred embodiment the organism according to the invention does not express a KDG kinase (kdgK) activity. It can either be an organism which naturally does not express kdgK or it can be an organism which naturally expresses a kdgK but in which the corresponding gene has been inactivated, e.g. by gene disruption or other suitable methods well-known to the person skilled in the art.

The present invention also relates to the use of an enzyme having keto acid decarboxylase activity or of a polynucleotide encoding such an enzyme in a method for converting KDG into 2-deoxy-D-ribose. With respect to the preferred embodiments the same applies as has already been set forth in connection with the method according to the present invention.

These and other embodiments are disclosed and encompassed by the description and examples of the present invention. The disclosure content of any references cited above or below is herewith incorporated into the present application. Further literature concerning any one of the methods, uses and compounds to be employed in accordance with the present invention may be retrieved from public libraries, using for example electronic devices. For example the public database "Medline" may be utilized which is available on the Internet, for example under <http://www.ncbi.nlm.nih.gov/PubMed/medline.html>. Further databases and addresses, such as <http://www.ncbi.nlm.nih.gov/>, <http://www.infobiogen.fr/>, http://www.fmi.ch/biology/research_tools.html, <http://www.tigr.org/>, are known to the person skilled in the art and can also be obtained using, e.g., <http://www.google.de>. An overview of patent information in biotechnology and a survey of relevant sources of patent information useful for retrospective searching and for current awareness is given in Berks, TIBTECH 12 (1994), 352-364.

Furthermore, the term "and/or" when occurring herein includes the meaning of "and", "or" and "all or any other combination of the elements connected by said term".

EXAMPLES

Example 1: Cloning of a gene encoding a D-gluconate dehydratase from *Agrobacterium tumefaciens* strain C58 (CIP 104333)

Agrobacterium tumefaciens strain C58 (CIP 104333) was obtained from Institut Pasteur Collection (CIP, Paris, France). Chromosomal DNA was extracted and a D-gluconate dehydratase gene was amplified by PCR according to standard protocols using the following primers:

5'-CCCTTAATTAATGACGACATCTGATAATCTTC-3', depicted in SEQ ID N° 5;

5'-TTTGC GGCCGCTTAGTGGTTATCGCGCGGC-3', depicted in SEQ ID N° 6;
5'-CCCGGTACCATGACGACATCTGATAATCTTC-3', depicted in SEQ ID N° 7;
A first DNA fragment amplified using the two primers depicted in SEQ ID N° 5 and SEQ ID N° 6, was ligated into a pUC18-derived vector previously digested by *PacI* and *NotI* to yield the plasmid pVDM80. A second DNA fragment amplified using the two primers depicted in SEQ ID N° 6 and SEQ ID N° 7, was ligated into a pET29a vector (Novagen) previously digested by *KpnI* and *NotI* to yield the plasmid pVDM82. The nucleotide sequence of the cloned gene is depicted in SEQ ID N° 1 and the sequence of the polypeptide encoded by this gene is depicted in SEQ ID N° 2.

Example 2: Expression of a D-gluconate dehydratase activity in Escherichia coli and preparation of 2-dehydro-3-deoxy-D-gluconate from D-gluconate

Competent cells of *E. coli* BL21 were transformed with the pVDM82 plasmid constructed as described in example 1 yielding strain +1289. Strain + 1289 was cultivated at 30°C in Luria-Bertani (LB) medium (Difco) containing 30 mg/l kanamycin until OD(600 nm) reached a value of 0.6. Then isopropyl- β -D-thiogalactopyranoside (IPTG) was added to a 0.5 mM final concentration. After a further cultivation period of 2 hours and 30 minutes, cells were collected by centrifugation and washed once with 20 mM sodium phosphate buffer pH 7.2. A cell extract was prepared by suspending about 5 g of cells in 10 ml of Tris-HCl 50 mM pH 8.5 buffer containing 10000 units lysozyme (Ready-Lyse, Epicentre, Madison, Wisconsin) and 1 mM EDTA, and incubating the suspension at 30°C for 15 minutes. Then 10000 kUnits deoxyribonuclease I (DNase I, Sigma) as well as 5 mM MgCl₂ were added to the preparation which was incubated at 30°C for an additional period of 15 minutes. The cell extract thus obtained was kept frozen at -20°C before use.

1.5 ml of the cell extract was mixed with 2M sodium or potassium D-gluconate in a total volume of 10 ml. This preparation was incubated at 37°C after the pH has been adjusted to 8.5. The progression of 2-dehydro-3-deoxy-D-gluconate (KDG) synthesis was followed by analysing aliquots taken after increasing periods of incubation. Several dilution parts of these aliquots were deposited on silica plates and chromatographed in the following solvent system: isopropanol / water (90/10). A yellow spot of KDG (*R_f* ~0.40) was detected after revelation with p-anisaldehyde.

KDG was also quantitated using a spectrophotometric assay based on the reaction with semicarbazide hydrochloride as described by Mac Gee (J. Biol. Chem. 1954, 210, 617-626). Typically, after a 30h period of incubation and using the spectrophotometric assay, KDG concentration ranged from 1.5 to 2 M.

The sodium or potassium 2-dehydro-3-deoxy-D-gluconate solution thus obtained could be used as such for further synthetic steps. 2-Dehydro-3-deoxy-D-gluconic acid could also be prepared from such a solution applying published protocols (Bender, Anal. Biochem. 1974, 61, 275-279). A crude preparation of a mixture of 2-dehydro-3-deoxy-D-gluconic acid and KCl could also be obtained by adding one equivalent of HCl to a potassium 2-dehydro-3-deoxy-D-gluconate solution which was then evaporated.

Example 3: Cloning of a gene encoding a D-glucosaminase from Agrobacterium tumefaciens strain C58 (CIP 104333)

Agrobacterium tumefaciens strain C58 (CIP 104333) was obtained from Institut Pasteur Collection (CIP, Paris, France). Chromosomal DNA was extracted and a D-glucosaminase gene was amplified by PCR according to standard protocols using the following primers:

5'-CCCTTAATTAATGCAGTCTTCTTCAGCTCTTC-3', depicted in SEQ ID N° 8;

5'-TTTGCGGCCCGCCTAGTGAAAGAAGGTTGTGTAGAT-3', depicted in SEQ ID N° 9;

5'-AAATCATGACTATGCAGTCTTCTTCAGCTCTTCG-3', depicted in SEQ ID N° 10;

5'-TATAGATCTCTAGTGAAAGAAGGTTGTGTAGAT-3', depicted in SEQ ID N° 11;

A first DNA fragment amplified using the two primers depicted in SEQ ID N° 8 and SEQ ID N° 9, was ligated into a pUC18-derived vector previously digested by *PacI* and *NotI* to yield the plasmid pKDGb1. A second DNA fragment amplified using the two primers depicted in SEQ ID N° 10 and SEQ ID N° 11, was ligated into a pQE60 vector (Qiagen) previously digested by *BspH1* and *BglII* to yield the plasmid pEP18. The nucleotide sequence of the cloned gene is depicted in SEQ ID N° 3 and the sequence of the polypeptide encoded by this gene is depicted in SEQ ID N° 4.

Example 4: Expression of a D-glucosaminase deaminase activity in Escherichia coli and preparation of 2-dehydro-3-deoxy-D-gluconic acid from D-glucosamine

Competent cells of E. coli MG1655 were transformed with the pEP18 plasmid constructed as described in example 1 and pREP4 (Qiagen) yielding strain +1068. Strain + 1068 was cultivated at 37°C in LB medium containing 30 mg/l kanamycin and 100 mg/l ampicillin until OD(600 nm) reached a value of 0.6. Then IPTG was added to a 0.5 mM final concentration. After a further cultivation period of 2 hours and 30 minutes, cells were collected by centrifugation and washed once with 20 mM sodium phosphate buffer pH 7.2. A cell extract was prepared using the protocol described in example 2.

2 ml of the cell extract was mixed with 100 mM sodium or potassium D-glucosamine and 0.1 mM pyridoxal phosphate in a total volume of 5 ml. This preparation was incubated at 37°C after the pH has been adjusted to 7.5. The progression of 2-dehydro-3-deoxy-D-gluconate (KDG) synthesis was followed using the protocols described in example 2. Typically, after a 30h period of incubation and using the spectrophotometric assay described in example 2, KDG concentration ranged from 50 to 100 mM.

Example 5: Preparation of 2-deoxy-D-ribonate from 2-dehydro-3-deoxy-D-gluconate

0.5 ml of a 31% hydrogen peroxide solution were added to 5 ml of a 1M potassium 2-dehydro-3-deoxy-D-gluconate (KDG) solution at 25°C. The progression of KDG decarboxylation was followed both by the observation of bubbles resulting from the release of carbon dioxide and by the disappearance of KDG using the thin layer chromatography protocol described in example 2. Typically, after a 3h period of reaction the concentration of residual KDG was less than 10 mM.

Example 6: Preparation of 2-deoxy-D-ribitol from 2-deoxy-D-ribonolactone

0.2 g of Rhodium (5 % on carbon) catalyst was added to an aqueous solution of 1 g 2-deoxy-D-ribonolactone prepared following a method described by Deriaz (J. Chem. Soc. (1949), 1879-1883) for the synthesis of 2-deoxy-L-ribonolactone. Hydrogenation of 2-deoxy-D-ribonolactone was performed at 130°C under a pressure of 80 bars. The solution obtained after filtration of the reaction mixture was evaporated. The residue was dissolved in ethyl acetate and further purified by chromatography on a silica column. The solvent was removed in vacuo leading to a yellow oil (yield 85%). The compound thus obtained was identical with 2-deoxy-D-ribitol obtained by reduction of 2-deoxy-D-ribose as described by Rabow (J. Am. Chem. Soc. 122 (1999), 3196-3203).

Example 7: Preparation of 1-N-morpholino-3,4,5-trihydroxypentene-1 from 2-dehydro-3-deoxy-D-gluconate

2 g of 2-dehydro-3-deoxy-D-gluconic acid were suspended in 150 ml benzene. 1.1 ml morpholine and 100 mg p-toluenesulfonic acid were added to the suspension and the reaction mixture was refluxed for 3 hours. Water formed by this reaction was removed by distillation. Benzene was decanted. Solid compounds attached to the vessel were collected, washed with acetone and dried. The main compound present in this preparation (yield 40%) was further purified by column chromatography on a silica column using a gradient of methanol in chloroform. Fractions containing 1-N-morpholino-3,4,5-trihydroxypentene-1 were pooled and solvent was removed in vacuo.

¹H-NMR (D₂O): δ = 3.15 ppm (4H, t, morpholine), 3.8 ppm (4H, t, morpholine), 3.4 to 4 ppm, (4H, m, 5a-H, 5b-H, 4-H, 3-H), 6.3 and 6.8 ppm (2H, 2d, 1-H and 2-H, J = 4 Hz).

Example 8: Cloning of a gene encoding a pyruvate decarboxylase from *Zymomonas mobilis*

Zymomonas mobilis strain B-806 (CIP 102538T) was obtained from Institut Pasteur Collection (CIP, Paris, France). Chromosomal DNA was extracted and a pyruvate

decarboxylase gene was amplified by PCR according to standard protocols using the following primers:

5'-GCGTTAATTAATGAGTTATACTGTCGGTACC-3', depicted in SEQ ID N° 12;

5'-TATGCGGCCGCTTAGAGGAGCTTGTTAACAGG-3', depicted in SEQ ID N° 13;

The DNA fragment amplified using the two primers depicted in SEQ ID N° 12 and SEQ ID N° 13, was ligated either into pSP100 or into pEVL5 (respectively a pUC18-derived or a pQE70-derived vector as described below) previously digested by *PacI* and *NotI* to yield respectively plasmid pEVL107 and plasmid pEVL420. The nucleotide sequence of the cloned gene as well as the encoded sequence of the corresponding polypeptide can be found at GenBank (accession number AF124349) and is shown in SEQ ID NO: 18 and SEQ ID NO: 19, respectively.

Plasmid pSP100 was obtained by introducing a ribosomal binding site, a *PacI* and a *NotI* restriction sites into a pUC18 vector previously digested by *EcoRI* and *BamHI* using standard protocols. The complete nucleotide sequence of pSP100 is depicted in SEQ ID N° 14.

Plasmid pEVL5 was obtained by introducing a ribosomal binding site, a *PacI* and a *NotI* restriction sites into a pQE70 vector (Qiagen) previously digested by *EcoRI* and *BamHI* using standard protocols. The complete nucleotide sequence of pEVL5 is depicted in SEQ ID N° 15.

Example 9: Cloning of a gene encoding a pyruvate decarboxylase from *Saccharomyces cerevisiae*

Chromosomal DNA was extracted from *Saccharomyces cerevisiae* strain S288C (ATCC 204508) and a pyruvate decarboxylase gene was amplified by PCR according to standard protocols using the following primers:

5'-ATATTTAATTAATGTCTGAAATTACTTTGG-3', depicted in SEQ ID N° 16;

5'-ATATGCGGCCGCTTATTGCTTAGCGTTGGT-3', depicted in SEQ ID N° 17;

The DNA fragment amplified using the two primers depicted in SEQ ID N° 16 and SEQ ID N° 17, was ligated either into pSP100 or into pEVL5 (respectively a pUC18-derived or a pQE70-derived vector as described in example 8) previously digested by

PacI and NotI to yield respectively plasmid pVDM61 and plasmid pEVL419. The nucleotide sequence of the cloned gene as well as the encoded sequence of the corresponding polypeptide can be found at GenBank (accession number NC001144) and is shown in SEQ ID NO: 20 and SEQ ID NO: 21, respectively.

Example 10: Expression of a pyruvate decarboxylase activity in Escherichia coli and enzymatic synthesis of 2-deoxy-D-ribose from 2-dehydro-3-deoxy-D-gluconate

Expression of pyruvate decarboxylase and preparation of cell-free extracts

Competent cells of *E. coli* MG1655 strain were transformed with either pEVL107 or pVDM61 (constructed as described in examples 8 and 9) yielding respectively strain +1735 and strain +844. These strains were cultivated at 37°C in Luria-Bertani (LB) medium (Difco) containing 100 mg/l ampicillin until OD(600 nm) reached a value around 1.5.

Competent cells of *E. coli* MG1655 strain harbouring pREP4 plasmid (Qiagen) were transformed with either pEVL420 or pEVL419 (constructed as described in Examples 8 and 9) yielding respectively strain +3150 and +3148. These strains were cultivated at 37°C in Luria-Bertani (LB) medium (Difco) containing 100 mg/l ampicillin and 30 mg/l kanamycin until OD(600 nm) reached a value of 0.6. Then isopropyl- β -D-thiogalactopyranoside (IPTG) was added to a 0.5 mM final concentration. After a further cultivation period of 2 hours and 30 minutes, cells were collected by centrifugation and washed once with 20 mM sodium phosphate buffer pH 7.2.

For each strain a cell-free extract was prepared using the same protocol as described in Example 2. Then crude cell-free extracts were passed through a PD-10 column (Amersham) equilibrated with 50 mM Tris-acetate buffer pH 6 and stored at -20°C.

Enzymatic synthesis of 2-deoxy-D-ribose from 2-dehydro-3-deoxy-D-gluconate

1.0 ml of cell-free extract was mixed with 20 mM sodium 2-dehydro-3-deoxy-D-gluconate, 0.5 mM thiamine pyrophosphate and 5 mM MgCl₂ in a total volume of 1.5 ml of 50 mM Tris-acetate buffer pH 6. The progression of 2-deoxy-D-ribose (DRI) synthesis was followed by analysing aliquots taken after increasing periods of

incubation at 37°C. About 1 µl of each aliquot which had been previously concentrated five-fold by evaporation was deposited on a silica plate and chromatographed in the following solvent system: butanol / triethylamine / water (10/2/5). A blue spot of DRI (Rf ~0.50) was detected after revelation with orcinol when using cell-free extracts of either strain +3150 or +3148 after a period of incubation of 65 hours. The crude preparation containing the spot corresponding to DRI was concentrated and passed through a 1.5 ml silica column equilibrated with isopropanol. The fractions containing the expected DRI compound were pooled, concentrated and the resulting sample analysed by mass spectrometry. The results of such an analysis confirmed the identity of the isolated compound with DRI, and the production of DRI from KDG catalysed by pyruvate decarboxylase either from *Zymomonas mobilis* or from *Saccharomyces cerevisiae*.

Example 11: Cloning of a gene encoding a pyruvate decarboxylase from *Acetobacter pasteurianus*, expression of encoded pyruvate decarboxylase activity in *Escherichia coli* and enzymatic synthesis of 2-deoxy-D-ribose from 2-dehydro-3-deoxy-D-gluconate

Acetobacter pasteurianus strain NCIB 8618 (DSMZ 2347) was obtained from DSMZ Collection (Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Braunschweig, Germany). Chromosomal DNA was extracted from the cells and a pyruvate decarboxylase gene was amplified by PCR according to standard protocols using the following primers:

5'-TCTTTAATTAATGGGTTGTCCGTCATTCATATA-3', depicted in SEQ ID N° 22;

5'-CTAAAGCTTTTAGGCCAGAGTGGTCTTGCGCG-3', depicted in SEQ ID N° 23;

The DNA fragment amplified using the two primers depicted in SEQ ID N° 22 and SEQ ID N° 23, was ligated either into pSP100 or into pEVL5 (respectively a pUC18-derived or a pQE70-derived vector as described in example 8) previously digested by *PacI* and *NotI* to yield respectively plasmid pEVL541 and plasmid pEVL560. The nucleotide sequence SEQ ID N° 24 of the cloned gene as well as the encoded sequence of the corresponding polypeptide SEQ ID N° 25 can be found at GenBank (accession number AF368435).

Competent cells of *E. coli* MG1655 strain were transformed with pEVL541 yielding strain +3559. Competent cells of *E. coli* MG1655 strain harbouring pREP4 plasmid (Qiagen) were transformed with pEVL560 yielding strain +3924. These strains were cultivated and cell-free extracts were prepared as described in Example 10. Cell-free extracts were incubated with KDG and the progression of 2-deoxy-D-ribose (DRI) synthesis was followed as described in Example 10. A spot corresponding to DRI was observed indicating that pyruvate decarboxylase from *Acetobacter pasteurianus* was able to decarboxylate KDG into DRI.

Example 12: Cloning of a gene encoding a pyruvate decarboxylase from *Zymobacter palmae*, expression of encoded pyruvate decarboxylase activity in *Escherichia coli* and enzymatic synthesis of 2-deoxy-D-ribose from 2-dehydro-3-deoxy-D-gluconate

Zymobacter palmae strain T109 (DSMZ10491) was obtained from DSMZ Collection (Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Braunschweig, Germany). Chromosomal DNA was extracted from the cells and a pyruvate decarboxylase gene was amplified by PCR according to standard protocols using the following primers:

5'-ATCTTAATTAATGTATACCGTTGGTATGTACT-3', depicted in SEQ ID N° 26;

5'-TATGCGGCCGCTTACGCTTGTGGTTTGCAGAGT-3', depicted in SEQ ID N° 27;

The DNA fragment amplified using the two primers depicted in SEQ ID N° 26 and SEQ ID N° 27, was ligated either into pSP100 or into pEVL5 (respectively a pUC18-derived or a pQE70-derived vector as described in example 8) previously digested by *PacI* and *NotI* to yield respectively plasmid pEVL546 and plasmid pEVL561. The nucleotide sequence of the cloned gene as well as the encoded sequence of the corresponding polypeptide is shown in SEQ ID NOs: 28 and 29, respectively and can be found at GenBank (accession number AF474145).

Competent cells of *E. coli* MG1655 strain were transformed with pEVL546 yielding strain +3568. Competent cells of *E. coli* MG1655 strain harbouring pREP4 plasmid (Qiagen) were transformed with pEVL560 yielding strain +3923. These strains were cultivated and cell-free extracts were prepared as described in Example 10. Cell-free

extracts were incubated with KDG and the progression of 2-deoxy-D-ribose (DRI) synthesis was followed as described in Example 10. A spot corresponding to DRI was observed indicating that pyruvate decarboxylase from *Zymobacter palmae* was able to decarboxylate KDG into DRI.

Example 13. Cloning of a gene encoding a benzoylformate decarboxylase from *Pseudomonas putida*, expression of encoded benzoylformate decarboxylase activity in *Escherichia coli* and enzymatic synthesis of 2-deoxy-D-ribose from 2-dehydro-3-deoxy-D-gluconate.

Pseudomonas putida strain Migula (DSMZ 291) was obtained from DSMZ Collection (Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Braunschweig, Germany). Chromosomal DNA was extracted from and a benzoylformate decarboxylase gene was amplified by PCR according to standard protocols using the following primers:

5'-CTATTAATTAATGGCTTCGGTACACGGCACCA-3', depicted in SEQ ID N° 30;

5'-TATGCGGCCGCTTACTTCACCGGGCTTACGGTGC-3', depicted in SEQ ID N° 31;

The DNA fragment amplified using the two primers depicted in SEQ ID N° 30 and SEQ ID N° 31, was ligated either into pSP100 or into pEVL5 (respectively a pUC18-derived or a pQE70-derived vector as described in example 8) previously digested by *PacI* and *NotI* to yield respectively plasmid pEVL681 and plasmid pEVL670. The nucleotide sequence SEQ ID N° 32 of the cloned gene as well as the encoded sequence of the corresponding polypeptide SEQ ID N° 33 can be found at GenBank (accessing number AY143338).

Competent cells of *E. coli* MG1655 strain were transformed with pEVL681 yielding strain +4050. Competent cells of *E. coli* MG1655 strain harbouring pREP4 plasmid (Qiagen) were transformed with pEVL670 yielding strain +3927. Those strains were cultivated and cell-free extracts were prepared as described in example 10. Cell-free extracts were incubated with KDG and the progression of 2-deoxy-D-ribose (DRI) synthesis was followed as described in example 10. A spot corresponding to DRI was observed indicating that benzoylformate decarboxylase from *Pseudomonas putida* was able to decarboxylate KDG into DRI.

Preparative enzymatic synthesis of 2-deoxy-D-ribose

100 µl of cell-free extract from strain +3927 (containing 2.5 mg of bacterial proteins) were mixed with 300 mM sodium 2-dehydro-3-deoxy-D-gluconate, 0.5 mM thiamine pyrophosphate and 5 mM MgCl₂ in a total volume of 0.5 ml of 80 mM potassium phosphate buffer pH 6. After a period of incubation of 16 and 40 hours, few µl of a solution of HCl 2N were added to the incubation mixture until the pH reached a value of 6. The progression of 2-deoxy-D-ribose (DRI) synthesis was also followed by analysing aliquots taken after increasing periods of incubation at 37°C. About 1 µl of each aliquot was deposited on a silica plate and chromatographed as described in example 10. The concentration of 2-deoxy-D-ribose was estimated to be about 200 mM by comparison with standard solutions. ¹³C NMR analysis of the crude mixture confirmed that the compound formed from 2-dehydro-3-deoxy-D-gluconate was 2-deoxy-D-ribose, and that the concentration of 2-deoxy-D-ribose was closed to 25 g/l. Another preparative enzymatic synthesis was performed in the same conditions except that no addition of acid was made along the incubation period. In those conditions, the concentration of 2-deoxy-D-ribose was closed to 10 g/l, far lower than the concentration reached in the preceding experiment for which the pH had been controlled and regularly adjusted to a value of 6.

Example 14. Enzymatic synthesis of 2-deoxy-D-ribose from D-gluconate

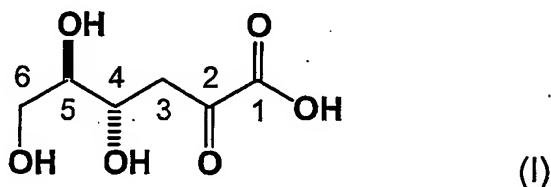
One pot enzymatic synthesis of 2-deoxy-D-ribose from D-gluconate was achieved as follows, using D-gluconate dehydratase encoded by gcnD gene of *Agrobacterium tumefaciens* and pyruvate decarboxylase from *Zymomonas mobilis*:

50 µl of cell-free extract from strain +1289 (containing 1.5 mg of bacterial proteins) and 400 µl of cell-free extract from strain +3150 (containing 17 mg of bacterial proteins after concentration by ultrafiltration) prepared as described respectively in example 2 and in example 10, were mixed with 50 mM potassium D-gluconate, 0.5 mM thiamine pyrophosphate and 5 mM MgCl₂ in a total volume of 0.5 ml of 50 mM N-(2-hydroxyethyl)piperazine-N'-(2-ethanesulfonic acid) (HEPES) buffer pH 7. The progression of 2-deoxy-D-ribose (DRI) synthesis was also followed by analysing

aliquots taken after increasing periods of incubation at 37°C. After a period of incubation of 18 hours, about 1 µl of the incubation mixture was deposited on a silica plate and chromatographed as described in example 10. The concentration of 2-deoxy-D-ribose was estimated to be about 1 g/l by comparison with standard solutions.

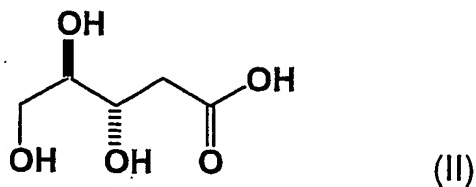
Claims

1. A method for producing 2'-deoxynucleosides or 2'-deoxynucleoside precursors from a compound of formula (I) or its salts



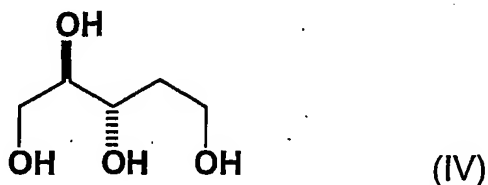
or a protected form thereof in a process comprising a decarboxylation step.

2. The method of claim 1 wherein the decarboxylation step cleaves the C1-C2 bond of the compound of formula (I) or its salts or a protected form thereof.
3. The method of claim 1 or 2, wherein the decarboxylation step is directly carried out on the compound of formula (I) or its salts or a protected form thereof.
4. The method of any of claims 1 to 3, wherein the decarboxylation step takes place by reacting the compound of formula (I) or its salts or a protected form thereof with hydrogen peroxide to yield a compound of formula (II) or its salts.



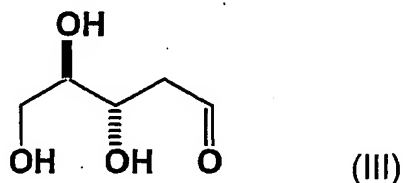
or a protected form thereof as a 2'-deoxynucleoside precursor.

5. The method of claim 4, further comprising the conversion of the compound of formula (II) or its salts or a protected form thereof into a compound of formula (IV)



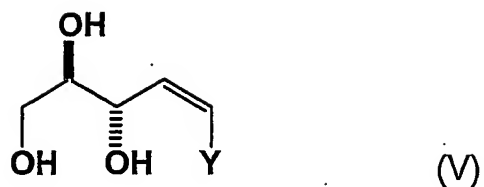
or a protected form thereof as a 2'-deoxynucleoside precursor.

6. The method of claim 4, further comprising the conversion of the compound of formula (II) or its salts or a protected form thereof into a compound of formula (III)



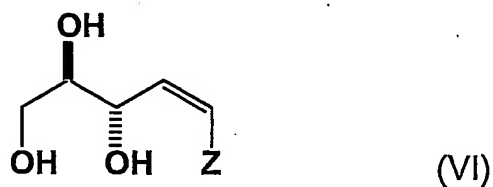
or a protected form thereof as a 2'-deoxynucleoside precursor.

7. The method of claim 6, comprising the conversion of the compound of formula (II) or its salts or a protected form thereof into the compound of formula (IV) or a protected form thereof as an intermediate which is then converted to the compound of formula (III) or a protected form thereof.
8. The method of any of claims 1 to 3, wherein the decarboxylation step takes place by reacting the compound of formula (I) or its salts or a protected form thereof with an amine Y-H, wherein H represents a hydrogen atom bound to the nitrogen atom of the amino group, to produce a compound of formula (V),



or its respective trans isomer or a protected form thereof, as a 2'-deoxynucleoside precursor.

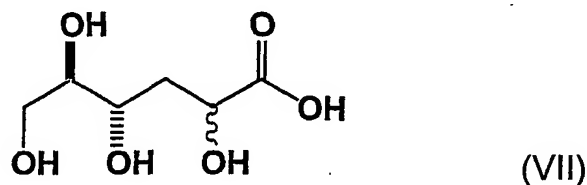
9. The method of claim 8, wherein Y-H represents a linear or cyclic secondary amine.
10. The method of claims 8 or 9, wherein Y-H is morpholine, pyrrolidine, piperidine, N-methyl piperazine or diethylamine.
11. The method of any of claims 8 to 10, further comprising the step of reacting a compound of formula (V) or its trans isomer or a protected form thereof with Z-H, wherein H represents a hydrogen atom and Z represents a leaving group, to produce a compound of formula (VI)



or its respective trans isomer or a protected form thereof, as a 2'-deoxynucleoside precursor.

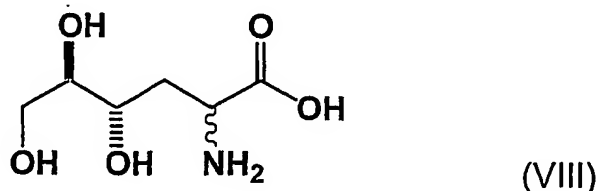
12. The method of claim 11, wherein Z-H is water, to produce a compound of formula (III) or a protected form thereof as a 2'-deoxynucleoside precursor.

13. The method of claim 1 or 2, wherein the compound of formula (I) or its salts or a protected form thereof is converted to a compound of formula (VII), or its salts or a protected form thereof or a mixture of the respective epimers,



which is then decarboxylated to yield a compound of formula (III) or a protected form thereof as a 2'-deoxynucleoside precursor.

14. The method of claim 13, wherein the conversion of (I) or its salts or a protected form thereof to (VII) or a protected form thereof takes place by reduction with sodium borohydride or by hydrogenation using Nickel Raney or Platinum oxide catalyst.
15. The method of claim 13 to 14, wherein the decarboxylation step takes place by reaction with hydrogen peroxide.
16. The method of claim 1 or 2, wherein the compound of formula (I) or its salts or a protected form thereof is converted to a compound of formula (VIII), or its salts or a protected form thereof or a mixture of the respective epimers,



which is then decarboxylated to yield a compound of formula (III) or a protected form thereof as a 2'-deoxynucleoside precursor.

17. The method of claim 16, wherein a compound of formula (VIII) or a protected form thereof or a mixture of the respective epimers is reacted with ninhydrin, thereby leading to the compound (III) or a protected form thereof.
18. The method of claim 16 or 17, wherein the conversion of (I) or its salts or a protected form thereof to (VIII) or a protected form thereof takes place by reductive amination with ammonia and sodium cyanoborohydride.
19. The method of any of claims 1 to 18, wherein the protective group(s) are independently chosen from acetate ester, benzoate ester, allyl ether, benzyl ether, trityl ether, ter-butyldimethylsilyl (TBDMS) ether, isopropylidene or a benzylidene acetal.
20. The method of any one of claims 1 to 3, wherein the decarboxylation step is effected by an enzymatic reaction comprising a single step.
21. The method of claim 20, wherein the enzymatic reaction is catalysed by an enzyme having keto acid decarboxylase activity.
22. The method of claim 21, wherein the enzyme having keto acid decarboxylase activity is a thiamine pyrophosphate (TPP) dependent keto acid decarboxylase.
23. The method of claim 22, wherein the TPP dependent keto acid decarboxylase is a pyruvate decarboxylase (EC 4.1.1.1), a benzoylformate decarboxylase (EC 4.1.1.7), an indolepyruvate decarboxylase (EC 4.1.1.74), a phosphonopyruvate decarboxylase, a sulfopyruvate decarboxylase (EC 4.1.1.79), an oxalyl-coenzyme A decarboxylase (EC 4.1.1.8), an oxoglutarate decarboxylase (EC 4.1.1.71) or a phenylpyruvate decarboxylase (EC 4.1.1.43).
24. The method of claim 23, wherein the pyruvate decarboxylase is of eukaryotic origin.

25. The method of claim 24, wherein the eukaryotic organism is a yeast organism.
26. The method of claim 25, wherein the yeast is *Saccharomyces cerevisiae*.
27. The method of claim 23, wherein the pyruvate decarboxylase is of prokaryotic origin.
28. The method of claim 27, wherein the prokaryotic organism is of the genus *Zymomonas*, *Zymobacter* or *Acetobacter*.
29. The method of claim 28, wherein the organism is of the species *Zymomonas mobilis*, *Zymobacter plamae* or *Acetobacter pasteurianus*.
30. The method of claim 23, wherein the benzoylformate decarboxylase is of prokaryotic origin.
31. The method of claim 30, wherein the prokaryotic organism is of the genus *Pseudomonas*.
32. The method of claim 31, wherein the organism is of the species *Pseudomonas putida*.
33. The method of any one of the claims 20 to 32, wherein the pH is regulated by addition of an acid between pH 5 and pH 9.
34. The method of claim 33, wherein the pH value is regulated between pH 6 and pH 8.
35. The method of claim 33 or 34, wherein the acid is HCl, H₂SO₄, D-gluconic acid or 2-dehydro-3-deoxy-D-gluconic acid.

36. The method of any one of claims 1 to 35, comprising the preliminary step of producing the compound of formula (I) from D-gluconate or a D-gluconate salt by the use of a gluconate dehydratase activity.
37. The method of claim 36, wherein the D-gluconate salt is potassium or sodium D-gluconate.
38. The method of claims 36 or 37, wherein the gluconate dehydratase is encoded by a polynucleotide comprising the nucleotide sequence selected from the group consisting of:
- (a) nucleotide sequences encoding a polypeptide comprising the amino acid sequence of SEQ ID N°2;
 - (b) nucleotide sequences comprising the coding sequence of SEQ ID N°1;
 - (c) nucleotide sequences encoding a fragment encoded by a nucleotide sequence of (a) or (b);
 - (d) nucleotide sequences hybridising with a nucleotide sequence of any one of (a) to (c); and
 - (e) nucleotide sequences which deviate from the nucleoside sequence of (d) as a result of degeneracy of the genetic code.
39. The method of any one of claims 1 to 35, comprising the preliminary step of producing the compound of formula (I) from D-glucosamine by the use of a glucosamine deaminase activity.
40. The method of claim 39, wherein the glucosamine deaminase is encoded by a polynucleotide comprising the nucleotide sequence selected from the group consisting of:
- (a) nucleotide sequences encoding a polypeptide comprising the amino acid sequence of SEQ ID N°4;
 - (b) nucleotide sequences comprising the coding sequence of SEQ ID N°3;
 - (c) nucleotide sequences encoding a fragment encoded by a nucleotide sequence of (a) or (b);

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- (d) nucleotide sequences hybridising with a nucleotide sequence of any one of (a) to (c); and
 - (e) nucleotide sequences which deviate from the nucleoside sequence of (d) as a result of degeneracy of the genetic code.
-
- 41. An organism which is capable of enzymatically converting D-gluconate into 2-dehydro-3-deoxy-D-gluconate due to the expression of a D-gluconate dehydratase and/or capable of enzymatically converting D-glucosamine into 2-dehydro-3-deoxy-D-gluconate due to the expression of a D-glucosamine deaminase and which is capable of enzymatically converting 2-dehydro-3-deoxy-D-gluconate by decarboxylation into 2-deoxy-D-ribose due to the expression of a keto acid decarboxylase.
 - 42. The organism of claim 41 which does not express a 2-dehydro-3-deoxy-D-gluconate kinase activity.
 - 43. The organism of claim 41 or 42 which does not express a 2-dehydro-3-deoxy-D-gluconate aldolase activity.
 - 44. The organism of any one of claims 41 to 43 which does not express a 2-deoxy-D-ribose aldolase activity.
 - 45. The method of any of claims 20 to 40 which is carried out by using an organism according to any one of claims 41 to 44.
 - 46. Use of a polynucleotide as defined in claim 38 or of a gluconate dehydratase encoded by such a polynucleotide in a method according to claims 36 or 37.
 - 47. Use of a polynucleotide as defined in claim 40 or of a glucosamine deaminase encoded by such a polynucleotide in a method according to claim 39.
 - 48. Use of an enzyme having keto acid decarboxylase activity or of a polynucleotide encoding such an enzyme in a method for converting a compound of the formula (I) into 2-deoxy-D-ribose.

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Marliere, Phillipe

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Glu Thr Gly Lys Arg Pro Thr Ala Gly Leu Asp Arg Asn Leu Ala Tyr			
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 545 550 555 560

gac cac cag acg ccg tgg cag gaa atc cag cgc ggt atc gtc agc cag 1728
 Asp His Gln Thr Pro Trp Gln Glu Ile Gln Arg Gly Ile Val Ser Gln
 565 570 575

atg gaa acc ggc gcg gtt ctg gaa ccg gcc gta aag tat cag cgc atc 1776
 Met Glu Thr Gly Ala Val Leu Glu Pro Ala Val Lys Tyr Gln Arg Ile
 580 585 590

gcc cag acc aag ggc ctg ccg cgc gat aac cac tga 1812
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<211> 603

<212> PRT

<213> Agrobacterium tumefaciens

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Leu Glu Arg Tyr Met Asn Phe Gly Leu Ser Gln Ala Glu Leu Gln Ser
 35 40 45

Asp Arg Pro Ile Ile Gly Ile Ala Gln Thr Gly Ser Asp Leu Ser Pro
 50 55 60

Cys Asn Arg His His Leu Glu Leu Ala Asn Arg Leu Arg Glu Gly Ile
 65 70 75 80

Arg Glu Ala Gly Gly Ile Ala Ile Glu Phe Pro Val His Pro Ile Gln
 85 90 95

Glu Thr Gly Lys Arg Pro Thr Ala Gly Leu Asp Arg Asn Leu Ala Tyr
 100 105 110

Leu Gly Leu Val Glu Val Leu Tyr Gly Tyr Pro Leu Asp Gly Val Val
 115 120 125

Leu Thr Ile Gly Cys Asp Lys Thr Thr Pro Ala Cys Leu Met Ala Ala
 130 135 140

Ala Thr Val Asn Ile Pro Ala Ile Ala Leu Ser Val Gly Pro Met Leu
 145 150 155 160

Asn Gly Trp Phe Arg Gly Glu Arg Thr Gly Ser Gly Thr Ile Val Trp
 165 170 175

Lys Ala Arg Glu Leu Leu Ala Lys Gly Glu Ile Asp Tyr Gln Gly Phe
 180 185 190

Val Lys Leu Val Ala Ser Ser Ala Pro Ser Thr Gly Tyr Cys Asn Thr
 195 200 205

Met Gly Thr Ala Thr Thr Met Asn Ser Leu Ala Glu Ala Leu Gly Met
 210 215 220

Gln Leu Pro Gly Ser Ala Ala Ile Pro Ala Pro Tyr Arg Asp Arg Gln
 225 230 235 240

Glu Val Ser Tyr Leu Thr Gly Leu Arg Ile Val Asp Met Val Arg Glu
 245 250 255

Asp Leu Lys Pro Ser Asp Ile Met Thr Lys Asp Ala Phe Ile Asn Ala
 260 265 270

Ile Arg Val Asn Ser Ala Ile Gly Gly Ser Thr Asn Ala Pro Ile His
 275 280 285

Leu Asn Gly Leu Ala Arg His Val Gly Val Glu Leu Thr Val Asp Asp
 290 295 300

Trp Gln Thr Tyr Gly Glu Asp Val Pro Leu Leu Val Asn Leu Gln Pro
 305 310 315 320

Ala Gly Glu Tyr Leu Gly Glu Asp Tyr Tyr His Ala Gly Gly Val Pro
 325 330 335

Ala Val Val Asn Gln Leu Met Thr Gln Gly Leu Ile Met Glu Asp Ala
 340 345 350

Met Thr Val Asn Gly Lys Thr Ile Gly Asp Asn Cys Arg Gly Ala Ile
 355 360 365

Ile Glu Asp Glu Lys Val Ile Arg Pro Tyr Glu Gln Pro Leu Lys Glu
 370 375 380

Arg Ala Gly Phe Arg Val Leu Arg Gly Asn Leu Phe Ser Ser Ala Ile
 385 390 395 400

Met Lys Thr Ser Val Ile Ser Glu Glu Phe Arg Gly Arg Tyr Leu Ser
 405 410 415

Asn Pro Asp Asp Pro Glu Ala Phe Glu Gly Arg Ala Val Val Phe Asp
 420 425 430

Gly Pro Glu Asp Tyr His His Arg Ile Asp Asp Pro Ser Leu Gly Ile
 435 440 445

Asp Ala Asn Thr Val Leu Phe Met Arg Gly Ala Gly Pro Ile Gly Tyr
 450 455 460

Pro Gly Ala Ala Glu Val Val Asn Met Arg Ala Pro Asp Tyr Leu Leu
 465 470 475 480

Lys Gln Gly Val Ser Ser Leu Pro Cys Ile Gly Asp Gly Arg Gln Ser
 485 490 495

Gly Thr Ser Gly Ser Pro Ser Ile Leu Asn Ala Ser Pro Glu Ala Ala
 500 505 510

Ala Gly Gly Gly Leu Ser Ile Leu Gln Thr Gly Asp Arg Val Arg Ile
 515 520 525

Asp Val Gly Arg Gly Lys Ala Asp Ile Leu Ile Ser Gly Glu Glu Leu
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Ala Lys Arg Tyr Glu Ala Leu Ala Ala Gln Gly Gly Tyr Lys Phe Pro
 545 550 555 560

Asp His Gln Thr Pro Trp Gln Glu Ile Gln Arg Gly Ile Val Ser Gln

565

570

575

Met Glu Thr Gly Ala Val Leu Glu Pro Ala Val Lys Tyr Gln Arg Ile
 580 585 590

Ala Gln Thr Lys Gly Leu Pro Arg Asp Asn His
 595 600

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<212> DNA

<213> Agrobacterium tumefaciens

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<222> (1)..(1269)

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tac cat gcc cag tcg aat atg atc ggc tct agc ccg gcg gac ggt ttg 96
 Tyr His Ala Gln Ser Asn Met Ile Gly Ser Ser Pro Ala Asp Gly Leu
 20 25 30

ctc gca ttg ccg ctt ctg acc gtc gat ctt gcc gtc tat cgc ggt aat 144
 Leu Ala Leu Pro Leu Leu Thr Val Asp Leu Ala Val Tyr Arg Gly Asn
 35 40 45

cgg gat cgc ttt ctt gcg ctt gtc tcg gcc cat gga gcg aag gcg gct 192
 Arg Asp Arg Phe Leu Ala Leu Val Ser Ala His Gly Ala Lys Ala Ala
 50 55 60

cca cat gcc aag acg ccg atg tgc ccg gag atc gcg atc gat ctg att 240
 Pro His Ala Lys Thr Pro Met Cys Pro Glu Ile Ala Ile Asp Leu Ile
 65 70 75 80

gaa gcc ggt gcc tgg ggc gcg acg gtc gcc gat ctc ttc cag gcg gaa 288
 Glu Ala Gly Ala Trp Gly Ala Thr Val Ala Asp Leu Phe Gln Ala Glu
 85 90 95

gtc ctg ctc aag gcc ggc gtg tcg aac ata ttg atc gcc aac cag atc 336
 Val Leu Leu Lys Ala Gly Val Ser Asn Ile Leu Ile Ala Asn Gln Ile
 100 105 110

ggc gga ttg aca tcc gcc aga cgc cta cgc atg ctc gca gat gct ttt Gly Gly Leu Thr Ser Ala Arg Arg Leu Arg Met Leu Ala Asp Ala Phe 115 120 125	384
ccg aaa gcc gag att atc tgc tgt gtc gat tct gtt cag gcc tcg gcc Pro Lys Ala Glu Ile Ile Cys Cys Val Asp Ser Val Gln Ala Ser Ala 130 135 140	432
aat ctg gtt cag gcc ttt caa ggg cgt gtg gat gcc cca ttc aag gtc Asn Leu Val Gln Ala Phe Gln Gly Arg Val Asp Ala Pro Phe Lys Val 145 150 155 160	480
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gca ctc gat gca aac atg gcg gcc ctg ttc gat ctc ctg acc gac agt Ala Leu Asp Ala Asn Met Ala Ala Leu Phe Asp Leu Leu Thr Asp Ser 210 215 220	672
ctt gca tcg ata cgc gaa aaa gat ccc ggg cgc ccg cta acg gtt tca Leu Ala Ser Ile Arg Glu Lys Asp Pro Gly Arg Pro Leu Thr Val Ser 225 230 235 240	720
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atc ttc ttc tct gat cac ggt gta tat cag cgc ggt ttc cag gca gtc Ile Phe Phe Ser Asp His Gly Val Tyr Gln Arg Gly Phe Gln Ala Val 275 280 285	864
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cct gag ccg ggg ctg gcg atc gtc ggc atg ggc atg ccg gat gta tcg Pro Glu Pro Gly Leu Ala Ile Val Gly Met Gly Met Arg Asp Val Ser 325 330 335	1008
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Leu Val Glu Ala Asp Leu Ser Ser Ser Ala Lys Val Gly Lys Leu Asn
      355      360      365

gac cag cat gcc ttc ttg tcc ttc ggg aac ggc agc agt ctg gca atc      1152
Asp Gln His Ala Phe Leu Ser Phe Gly Asn Gly Ser Ser Leu Ala Ile
      370      375      380

ggc gat gtc ata gaa ttc ggc atc tcg cat ccc tgc acg tgc ttc gat      1200
Gly Asp Val Ile Glu Phe Gly Ile Ser His Pro Cys Thr Cys Phe Asp
      385      390      395      400

cgc tgg cgc gtc ttt cac gga atc gat gga tca ggc cgg atc cag cgc      1248
Arg Trp Arg Val Phe His Gly Ile Asp Gly Ser Gly Arg Ile Gln Arg
      405      410      415

atc tac aca acc ttc ttt cac tag      1272
Ile Tyr Thr Thr Phe Phe His
      420

<210> 4
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<212> PRT
<213> Agrobacterium tumefaciens

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Leu Ala Leu Pro Leu Leu Thr Val Asp Leu Ala Val Tyr Arg Gly Asn
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Arg Asp Arg Phe Leu Ala Leu Val Ser Ala His Gly Ala Lys Ala Ala
50          55          60

Pro His Ala Lys Thr Pro Met Cys Pro Glu Ile Ala Ile Asp Leu Ile
65          70          75          80

Glu Ala Gly Ala Trp Gly Ala Thr Val Ala Asp Leu Phe Gln Ala Glu
85          90          95

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Val Leu Leu Lys Ala Gly Val Ser Asn Ile Leu Ile Ala Asn Gln Ile
 100 105 110

Gly Gly Leu Thr Ser Ala Arg Arg Leu Arg Met Leu Ala Asp Ala Phe
 115 120 125

Pro Lys Ala Glu Ile Ile Cys Cys Val Asp Ser Val Gln Ala Ser Ala
 130 135 140

Asn Leu Val Gln Ala Phe Gln Gly Arg Val Asp Ala Pro Phe Lys Val
 145 150 155 160

Phe Ile Glu Val Gly Val Gly Arg Thr Gly Ala Arg Thr Leu Asn Val
 165 170 175

Ala Lys Asp Ile Ile Asp Thr Ile Ser Thr Ser Ala Glu Ile Val Leu
 180 185 190

Ala Gly Val Ser Thr Tyr Glu Gly Ser Val Ser Gly Glu Thr Ser Glu
 195 200 205

Ala Leu Asp Ala Asn Met Ala Ala Leu Phe Asp Leu Leu Thr Asp Ser
 210 215 220

Leu Ala Ser Ile Arg Glu Lys Asp Pro Gly Arg Pro Leu Thr Val Ser
 225 230 235 240

Ala Gly Gly Ser Ile His Phe Asp Arg Val Leu Ala Ala Leu Val Pro
 245 250 255

Val Cys Glu Ala Asp Gly Asn Ala Thr Leu Leu Leu Arg Ser Gly Ala
 260 265 270

Ile Phe Phe Ser Asp His Gly Val Tyr Gln Arg Gly Phe Gln Ala Val
 275 280 285

Asp Ala Arg Asn Leu Leu Ala Ser Gly Lys Val Val Phe Lys Ala Ser
 290 295 300

Glu Ala Phe Gln Pro Ser Met Arg Ile Trp Ala Glu Val Ile Ser Val
 305 310 315 320

Pro Glu Pro Gly Leu Ala Ile Val Gly Met Gly Met Arg Asp Val Ser
 325 330 335

Phe Asp Gln Asp Leu Pro Val Ala Leu Arg Leu His Arg Asp Gly His
 340 345 350

Leu Val Glu Ala Asp Leu Ser Ser Ser Ala Lys Val Gly Lys Leu Asn
 355 360 365

Asp Gln His Ala Phe Leu Ser Phe Gly Asn Gly Ser Ser Leu Ala Ile
 370 375 380

Gly Asp Val Ile Glu Phe Gly Ile Ser His Pro Cys Thr Cys Phe Asp
 385 390 395 400

Arg Trp Arg Val Phe His Gly Ile Asp Gly Ser Gly Arg Ile Gln Arg
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Ile Tyr Thr Thr Phe Phe His
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<213> artificial sequence

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<210> 7

<211> 31

<212> DNA

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<220>

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<400> 7

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32

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<211> 35

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<400> 9

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<210> 10

<211> 34

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34

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<211> 33

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33

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<211> 31

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<210> 13

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<211> 2665

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<210> 15

<211> 3433

<212> DNA

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3433

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<211> 30

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<213> artificial sequence

<220>

<223> artificial sequence

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<210> 17

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<212> DNA

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<210> 18

<211> 1707

<212> DNA

<213> *Zymomonas mobilis*

<220>

<221> CDS

<222> (1)..(1707)

<223>

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<400> 18
atg agt tat act gtc ggt acc tat tta gcg gag cgg ctt gtc cag att      48
Met Ser Tyr Thr Val Gly Thr Tyr Leu Ala Glu Arg Leu Val Gln Ile
1          5          10          15

ggt ctc aag cat cac ttc gca gtc gcg ggc gac tac aac ctc gtc ctt      96
Gly Leu Lys His His Phe Ala Val Ala Gly Asp Tyr Asn Leu Val Leu
          20          25          30

ctt gac aac ctg ctt ttg aac aaa aac atg gag cag gtt tat tgc tgt      144
Leu Asp Asn Leu Leu Leu Asn Lys Asn Met Glu Gln Val Tyr Cys Cys
          35          40          45

aac gaa ctg aac tgc ggt ttc agt gca gaa ggt tat gct cgt gcc aaa      192
Asn Glu Leu Asn Cys Gly Phe Ser Ala Glu Gly Tyr Ala Arg Ala Lys
          50          55          60

ggc gca gca gca gcc gtc gtt acc tac agc gtc ggt gcg ctt tcc gca      240
Gly Ala Ala Ala Ala Val Val Thr Tyr Ser Val Gly Ala Leu Ser Ala
65          70          75          80

ttt gat gct atc ggt ggc gcc tat gca gaa aac ctt ccg gtt atc ctg      288
Phe Asp Ala Ile Gly Gly Ala Tyr Ala Glu Asn Leu Pro Val Ile Leu
          85          90          95

atc tcc ggt gct ccg aac aac aat gat cac gct gct ggt cac gtg ttg      336
Ile Ser Gly Ala Pro Asn Asn Asn Asp His Ala Ala Gly His Val Leu
          100          105          110

cat cac gct ctt ggc aaa acc gac tat cac tat cag ttg gaa atg gcc      384
His His Ala Leu Gly Lys Thr Asp Tyr His Tyr Gln Leu Glu Met Ala
          115          120          125

aag aac atc acg gcc gcc gct gaa gcg att tac acc ccg gaa gaa gct      432
Lys Asn Ile Thr Ala Ala Ala Glu Ala Ile Tyr Thr Pro Glu Glu Ala
          130          135          140

ccg gct aaa atc gat cac gtg att aaa act gct ctt cgt gag aag aag      480
Pro Ala Lys Ile Asp His Val Ile Lys Thr Ala Leu Arg Glu Lys Lys
          145          150          155          160

ccg gtt tat ctc gaa atc gct tgc aac att gct tcc atg ccc tgc gcc      528
Pro Val Tyr Leu Glu Ile Ala Cys Asn Ile Ala Ser Met Pro Cys Ala
          165          170          175

gct cct gga ccg gca agc gca ttg ttc aat gac gaa gcc agc gac gaa      576
Ala Pro Gly Pro Ala Ser Ala Leu Phe Asn Asp Glu Ala Ser Asp Glu
          180          185          190

gct tct ttg aat gca gcg gtt gaa gaa acc ctg aaa ttc atc gcc aac      624
Ala Ser Leu Asn Ala Ala Val Glu Glu Thr Leu Lys Phe Ile Ala Asn
          195          200          205

cgc gac aaa gtt gcc gtc ctc gtc ggc agc aag ctg cgc gca gct ggt      672
Arg Asp Lys Val Ala Val Leu Val Gly Ser Lys Leu Arg Ala Ala Gly
          210          215          220

gct gaa gaa gct gct gtc aaa ttt gct gat gct ctc ggt ggc gca gtt      720

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atc aat aac tat ggt tac acc atc gaa gtt atg atc cat gat ggt ccg 1440
 Ile Asn Asn Tyr Gly Tyr Thr Ile Glu Val Met Ile His Asp Gly Pro
 465 470 475 480
 tac aac aac atc aag aac tgg gat tat gcc ggt ctg atg gaa gtg ttc 1488
 Tyr Asn Asn Ile Lys Asn Trp Asp Tyr Ala Gly Leu Met Glu Val Phe
 485 490 495
 aac ggt aac ggt ggt tat gac agc ggt gct ggt aaa ggc ctg aag gct 1536
 Asn Gly Asn Gly Gly Tyr Asp Ser Gly Ala Gly Lys Gly Leu Lys Ala
 500 505 510
 aaa acc ggt ggc gaa ctg gca gaa gct atc aag gtt gct ctg gca aac 1584
 Lys Thr Gly Gly Glu Leu Ala Glu Ala Ile Lys Val Ala Leu Ala Asn
 515 520 525
 acc gac ggc cca acc ctg atc gaa tgc ttc atc ggt cgt gaa gac tgc 1632
 Thr Asp Gly Pro Thr Leu Ile Glu Cys Phe Ile Gly Arg Glu Asp Cys
 530 535 540
 act gaa gaa ttg gtc aaa tgg ggt aag cgc gtt gct gcc gcc aac agc 1680
 Thr Glu Glu Leu Val Lys Trp Gly Lys Arg Val Ala Ala Ala Asn Ser
 545 550 555 560
 cgt aag cct gtt aac aag ctc ctc tag 1707
 Arg Lys Pro Val Asn Lys Leu Leu
 565

<210> 19

<211> 568

<212> PRT

<213> *Zymomonas mobilis*

<400> 19

Met Ser Tyr Thr Val Gly Thr Tyr Leu Ala Glu Arg Leu Val Gln Ile
 1 5 10 15
 Gly Leu Lys His His Phe Ala Val Ala Gly Asp Tyr Asn Leu Val Leu
 20 25 30
 Leu Asp Asn Leu Leu Leu Asn Lys Asn Met Glu Gln Val Tyr Cys Cys
 35 40 45
 Asn Glu Leu Asn Cys Gly Phe Ser Ala Glu Gly Tyr Ala Arg Ala Lys
 50 55 60
 Gly Ala Ala Ala Ala Val Val Thr Tyr Ser Val Gly Ala Leu Ser Ala
 65 70 75 80

Phe Asp Ala Ile Gly Gly Ala Tyr Ala Glu Asn Leu Pro Val Ile Leu
 85 90 95

Ile Ser Gly Ala Pro Asn Asn Asn Asp His Ala Ala Gly His Val Leu
 100 105 110

His His Ala Leu Gly Lys Thr Asp Tyr His Tyr Gln Leu Glu Met Ala
 115 120 125

Lys Asn Ile Thr Ala Ala Ala Glu Ala Ile Tyr Thr Pro Glu Glu Ala
 130 135 140

Pro Ala Lys Ile Asp His Val Ile Lys Thr Ala Leu Arg Glu Lys Lys
 145 150 155 160

Pro Val Tyr Leu Glu Ile Ala Cys Asn Ile Ala Ser Met Pro Cys Ala
 165 170 175

Ala Pro Gly Pro Ala Ser Ala Leu Phe Asn Asp Glu Ala Ser Asp Glu
 180 185 190

Ala Ser Leu Asn Ala Ala Val Glu Glu Thr Leu Lys Phe Ile Ala Asn
 195 200 205

Arg Asp Lys Val Ala Val Leu Val Gly Ser Lys Leu Arg Ala Ala Gly
 210 215 220

Ala Glu Glu Ala Ala Val Lys Phe Ala Asp Ala Leu Gly Gly Ala Val
 225 230 235 240

Ala Thr Met Ala Ala Ala Lys Ser Phe Phe Pro Glu Glu Asn Pro His
 245 250 255

Tyr Ile Gly Thr Ser Trp Gly Glu Val Ser Tyr Pro Gly Val Glu Lys
 260 265 270

Thr Met Lys Glu Ala Asp Ala Val Ile Ala Leu Ala Pro Val Phe Asn
 275 280 285

Asp Tyr Ser Thr Thr Gly Trp Thr Asp Ile Pro Asp Pro Lys Lys Leu
 290 295 300

Val Leu Ala Glu Pro Arg Ser Val Val Val Asn Gly Ile Arg Phe Pro

305	310	315	320
Ser Val His Leu Lys Asp Tyr Leu Thr Arg Leu Ala Gln Lys Val Ser	325	330	335
Lys Lys Thr Gly Ala Leu Asp Phe Phe Lys Ser Leu Asn Ala Gly Glu	340	345	350
Leu Lys Lys Ala Ala Pro Ala Asp Pro Ser Ala Pro Leu Val Asn Ala	355	360	365
Glu Ile Ala Arg Gln Val Glu Ala Leu Leu Thr Pro Asn Thr Thr Val	370	375	380
Ile Ala Glu Thr Gly Asp Ser Trp Phe Asn Ala Gln Arg Met Lys Leu	385	390	395
Pro Asn Gly Ala Arg Val Glu Tyr Glu Met Gln Trp Gly His Ile Gly	405	410	415
Trp Ser Val Pro Ala Ala Phe Gly Tyr Ala Val Gly Ala Pro Glu Arg	420	425	430
Arg Asn Ile Leu Met Val Gly Asp Gly Ser Phe Gln Leu Thr Ala Gln	435	440	445
Glu Val Ala Gln Met Val Arg Leu Lys Leu Pro Val Ile Ile Phe Leu	450	455	460
Ile Asn Asn Tyr Gly Tyr Thr Ile Glu Val Met Ile His Asp Gly Pro	465	470	475
Tyr Asn Asn Ile Lys Asn Trp Asp Tyr Ala Gly Leu Met Glu Val Phe	485	490	495
Asn Gly Asn Gly Gly Tyr Asp Ser Gly Ala Gly Lys Gly Leu Lys Ala	500	505	510
Lys Thr Gly Gly Glu Leu Ala Glu Ala Ile Lys Val Ala Leu Ala Asn	515	520	525
Thr Asp Gly Pro Thr Leu Ile Glu Cys Phe Ile Gly Arg Glu Asp Cys	530	535	540

Thr Glu Glu Leu Val Lys Trp Gly Lys Arg Val Ala Ala Ala Asn Ser
 545 550 555 560

Arg Lys Pro Val Asn Lys Leu Leu
 565

<210> 20

<211> 1692

<212> DNA

<213> *Saccharomyces cerevisiae*

<220>

<221> CDS

<222> (1)..(1692)

<223>

<400> 20
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 Met Ser Glu Ile Thr Leu Gly Lys Tyr Leu Phe Glu Arg Leu Lys Gln
 1 5 10 15
 gtc aac gtt aac acc gtt ttc ggt ttg cca ggt gac ttc aac ttg tcc 96
 Val Asn Val Asn Thr Val Phe Gly Leu Pro Gly Asp Phe Asn Leu Ser
 20 25 30
 ttg ttg gac aag atc tac gaa gtt gaa ggt atg aga tgg gct ggt aac 144
 Leu Leu Asp Lys Ile Tyr Glu Val Glu Gly Met Arg Trp Ala Gly Asn
 35 40 45
 gcc aac gaa ttg aac gct gct tac gcc gct gat ggt tac gct cgt atc 192
 Ala Asn Glu Leu Asn Ala Ala Tyr Ala Ala Asp Gly Tyr Ala Arg Ile
 50 55 60
 aag ggt atg tct tgt atc atc acc acc ttc ggt gtc ggt gaa ttg tct 240
 Lys Gly Met Ser Cys Ile Ile Thr Thr Phe Gly Val Gly Glu Leu Ser
 65 70 75 80
 gct ttg aac ggt att gcc ggt tct tac gct gaa cac gtc ggt gtt ttg 288
 Ala Leu Asn Gly Ile Ala Gly Ser Tyr Ala Glu His Val Gly Val Leu
 85 90 95
 cac gtt gtt ggt gtc cca tcc atc tct gct caa gct aag caa ttg ttg 336
 His Val Val Gly Val Pro Ser Ile Ser Ala Gln Ala Lys Gln Leu Leu
 100 105 110
 ttg cac cac acc ttg ggt aac ggt gac ttc act gtt ttc cac aga atg 384
 Leu His His Thr Leu Gly Asn Gly Asp Phe Thr Val Phe His Arg Met

115	120	125	
tct gcc aac att tct gaa acc act gct atg atc act gac att gct acc			432
Ser Ala Asn Ile Ser Glu Thr Thr Ala Met Ile Thr Asp Ile Ala Thr			
130	135	140	
gcc cca gct gaa att gac aga tgt atc aga acc act tac gtc acc caa			480
Ala Pro Ala Glu Ile Asp Arg Cys Ile Arg Thr Thr Tyr Val Thr Gln			
145	150	155 160	
aga cca gtc tac tta ggt ttg cca gct aac ttg gtc gac ttg aac gtc			528
Arg Pro Val Tyr Leu Gly Leu Pro Ala Asn Leu Val Asp Leu Asn Val			
165	170	175	
cca gct aag ttg ttg caa act cca att gac atg tct ttg aag cca aac			576
Pro Ala Lys Leu Leu Gln Thr Pro Ile Asp Met Ser Leu Lys Pro Asn			
180	185	190	
gat gct gaa tcc gaa aag gaa gtc att gac acc atc ttg gct ttg gtc			624
Asp Ala Glu Ser Glu Lys Glu Val Ile Asp Thr Ile Leu Ala Leu Val			
195	200	205	
aag gat gct aag aac cca gtt atc ttg gct gat gct tgt tgt tcc aga			672
Lys Asp Ala Lys Asn Pro Val Ile Leu Ala Asp Ala Cys Cys Ser Arg			
210	215	220	
cac gac gtc aag gct gaa act aag aag ttg att gac ttg act caa ttc			720
His Asp Val Lys Ala Glu Thr Lys Lys Leu Ile Asp Leu Thr Gln Phe			
225	230	235 240	
cca gct ttc gtc acc cca atg ggt aag ggt tcc att gac gaa caa cac			768
Pro Ala Phe Val Thr Pro Met Gly Lys Gly Ser Ile Asp Glu Gln His			
245	250	255	
cca aga tac ggt ggt gtt tac gtc ggt acc ttg tcc aag cca gaa gtt			816
Pro Arg Tyr Gly Gly Val Tyr Val Gly Thr Leu Ser Lys Pro Glu Val			
260	265	270	
aag gaa gcc gtt gaa tct gct gac ttg att ttg tct gtc ggt gct ttg			864
Lys Glu Ala Val Glu Ser Ala Asp Leu Ile Leu Ser Val Gly Ala Leu			
275	280	285	
ttg tct gat ttc aac acc ggt tct ttc tct tac tct tac aag acc aag			912
Leu Ser Asp Phe Asn Thr Gly Ser Phe Ser Tyr Ser Tyr Lys Thr Lys			
290	295	300	
aac att gtc gaa ttc cac tcc gac cac atg aag atc aga aac gcc act			960
Asn Ile Val Glu Phe His Ser Asp His Met Lys Ile Arg Asn Ala Thr			
305	310	315 320	
ttc cca ggt gtc caa atg aaa ttc gtt ttg caa aag ttg ttg acc act			1008
Phe Pro Gly Val Gln Met Lys Phe Val Leu Gln Lys Leu Leu Thr Thr			
325	330	335	
att gct gac gcc gct aag ggt tac aag cca gtt gct gtc cca gct aga			1056
Ile Ala Asp Ala Ala Lys Gly Tyr Lys Pro Val Ala Val Pro Ala Arg			
340	345	350	
act cca gct aac gct gct gtc cca gct tct acc cca ttg aag caa gaa			1104

Thr	Pro	Ala	Asn	Ala	Ala	Val	Pro	Ala	Ser	Thr	Pro	Leu	Lys	Gln	Glu		
		355					360					365					
ttg	atg	ttg	aac	caa	ttg	ggt	aac	ttc	ttg	caa	gaa	ggt	gat	gtt	gtc	1152	
Trp	Met	Trp	Asn	Gln	Leu	Gly	Asn	Phe	Leu	Gln	Glu	Gly	Asp	Val	Val		
	370					375					380						
att	gct	gaa	acc	ggt	acc	tcc	gct	ttc	ggt	atc	aac	caa	acc	act	ttc	1200	
Ile	Ala	Glu	Thr	Gly	Thr	Ser	Ala	Phe	Gly	Ile	Asn	Gln	Thr	Thr	Phe		
	385				390					395					400		
cca	aac	aac	acc	tac	ggt	atc	tct	caa	gtc	tta	ttg	ggt	tcc	att	ggt	1248	
Pro	Asn	Asn	Thr	Tyr	Gly	Ile	Ser	Gln	Val	Leu	Trp	Gly	Ser	Ile	Gly		
			405						410					415			
ttc	acc	act	ggt	gct	acc	ttg	ggt	gct	gct	ttc	gct	gct	gaa	gaa	att	1296	
Phe	Thr	Thr	Gly	Ala	Thr	Leu	Gly	Ala	Ala	Phe	Ala	Ala	Glu	Glu	Ile		
			420					425					430				
gat	cca	aag	aag	aga	gtt	atc	tta	ttc	att	ggt	gac	ggt	tct	ttg	caa	1344	
Asp	Pro	Lys	Lys	Arg	Val	Ile	Leu	Phe	Ile	Gly	Asp	Gly	Ser	Leu	Gln		
		435					440					445					
ttg	act	gtt	caa	gaa	atc	tcc	acc	atg	atc	aga	ttg	ggc	ttg	aag	cca	1392	
Leu	Thr	Val	Gln	Glu	Ile	Ser	Thr	Met	Ile	Arg	Trp	Gly	Leu	Lys	Pro		
	450					455					460						
tac	ttg	ttc	gtc	ttg	aac	aac	gat	ggt	tac	acc	att	gaa	aag	ttg	att	1440	
Tyr	Leu	Phe	Val	Leu	Asn	Asn	Asp	Gly	Tyr	Thr	Ile	Glu	Lys	Leu	Ile		
	465				470				475					480			
cac	ggt	cca	aag	gct	caa	tac	aac	gaa	att	caa	ggt	ttg	gac	cac	cta	1488	
His	Gly	Pro	Lys	Ala	Gln	Tyr	Asn	Glu	Ile	Gln	Gly	Trp	Asp	His	Leu		
			485					490						495			
tcc	ttg	ttg	cca	act	ttc	ggt	gct	aag	gac	tat	gaa	acc	cac	aga	gtc	1536	
Ser	Leu	Leu	Pro	Thr	Phe	Gly	Ala	Lys	Asp	Tyr	Glu	Thr	His	Arg	Val		
			500					505					510				
gct	acc	acc	ggt	gaa	ttg	gac	aag	ttg	acc	caa	gac	aag	tct	ttc	aac	1584	
Ala	Thr	Thr	Gly	Glu	Trp	Asp	Lys	Leu	Thr	Gln	Asp	Lys	Ser	Phe	Asn		
		515					520					525					
gac	aac	tct	aag	atc	aga	atg	att	gaa	atc	atg	ttg	cca	gtc	ttc	gat	1632	
Asp	Asn	Ser	Lys	Ile	Arg	Met	Ile	Glu	Ile	Met	Leu	Pro	Val	Phe	Asp		
	530					535					540						
gct	cca	caa	aac	ttg	gtt	gaa	caa	gct	aag	ttg	act	gct	gct	acc	aac	1680	
Ala	Pro	Gln	Asn	Leu	Val	Glu	Gln	Ala	Lys	Leu	Thr	Ala	Ala	Thr	Asn		
	545				550					555					560		
gct	aag	caa	taa													1692	
Ala	Lys	Gln															

<210> 21

<211> 563

<212> PRT

<213> *Saccharomyces cerevisiae*

<400> 21

Met Ser Glu Ile Thr Leu Gly Lys Tyr Leu Phe Glu Arg Leu Lys Gln
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Val Asn Val Asn Thr Val Phe Gly Leu Pro Gly Asp Phe Asn Leu Ser
 20 25 30

Leu Leu Asp Lys Ile Tyr Glu Val Glu Gly Met Arg Trp Ala Gly Asn
 35 40 45

Ala Asn Glu Leu Asn Ala Ala Tyr Ala Ala Asp Gly Tyr Ala Arg Ile
 50 55 60

Lys Gly Met Ser Cys Ile Ile Thr Thr Phe Gly Val Gly Glu Leu Ser
 65 70 75 80

Ala Leu Asn Gly Ile Ala Gly Ser Tyr Ala Glu His Val Gly Val Leu
 85 90 95

His Val Val Gly Val Pro Ser Ile Ser Ala Gln Ala Lys Gln Leu Leu
 100 105 110

Leu His His Thr Leu Gly Asn Gly Asp Phe Thr Val Phe His Arg Met
 115 120 125

Ser Ala Asn Ile Ser Glu Thr Thr Ala Met Ile Thr Asp Ile Ala Thr
 130 135 140

Ala Pro Ala Glu Ile Asp Arg Cys Ile Arg Thr Thr Tyr Val Thr Gln
 145 150 155 160

Arg Pro Val Tyr Leu Gly Leu Pro Ala Asn Leu Val Asp Leu Asn Val
 165 170 175

Pro Ala Lys Leu Leu Gln Thr Pro Ile Asp Met Ser Leu Lys Pro Asn
 180 185 190

Asp Ala Glu Ser Glu Lys Glu Val Ile Asp Thr Ile Leu Ala Leu Val
 195 200 205

Lys Asp Ala Lys Asn Pro Val Ile Leu Ala Asp Ala Cys Cys Ser Arg
 210 215 220
 His Asp Val Lys Ala Glu Thr Lys Lys Leu Ile Asp Leu Thr Gln Phe
 225 230 235 240
 Pro Ala Phe Val Thr Pro Met Gly Lys Gly Ser Ile Asp Glu Gln His
 245 250 255
 Pro Arg Tyr Gly Gly Val Tyr Val Gly Thr Leu Ser Lys Pro Glu Val
 260 265 270
 Lys Glu Ala Val Glu Ser Ala Asp Leu Ile Leu Ser Val Gly Ala Leu
 275 280 285
 Leu Ser Asp Phe Asn Thr Gly Ser Phe Ser Tyr Ser Tyr Lys Thr Lys
 290 295 300
 Asn Ile Val Glu Phe His Ser Asp His Met Lys Ile Arg Asn Ala Thr
 305 310 315 320
 Phe Pro Gly Val Gln Met Lys Phe Val Leu Gln Lys Leu Leu Thr Thr
 325 330 335
 Ile Ala Asp Ala Ala Lys Gly Tyr Lys Pro Val Ala Val Pro Ala Arg
 340 345 350
 Thr Pro Ala Asn Ala Ala Val Pro Ala Ser Thr Pro Leu Lys Gln Glu
 355 360 365
 Trp Met Trp Asn Gln Leu Gly Asn Phe Leu Gln Glu Gly Asp Val Val
 370 375 380
 Ile Ala Glu Thr Gly Thr Ser Ala Phe Gly Ile Asn Gln Thr Thr Phe
 385 390 395 400
 Pro Asn Asn Thr Tyr Gly Ile Ser Gln Val Leu Trp Gly Ser Ile Gly
 405 410 415
 Phe Thr Thr Gly Ala Thr Leu Gly Ala Ala Phe Ala Ala Glu Glu Ile
 420 425 430
 Asp Pro Lys Lys Arg Val Ile Leu Phe Ile Gly Asp Gly Ser Leu Gln

435

440

445

Leu Thr Val Gln Glu Ile Ser Thr Met Ile Arg Trp Gly Leu Lys Pro
 450 455 460

Tyr Leu Phe Val Leu Asn Asn Asp Gly Tyr Thr Ile Glu Lys Leu Ile
 465 470 475 480

His Gly Pro Lys Ala Gln Tyr Asn Glu Ile Gln Gly Trp Asp His Leu
 485 490 495

Ser Leu Leu Pro Thr Phe Gly Ala Lys Asp Tyr Glu Thr His Arg Val
 500 505 510

Ala Thr Thr Gly Glu Trp Asp Lys Leu Thr Gln Asp Lys Ser Phe Asn
 515 520 525

Asp Asn Ser Lys Ile Arg Met Ile Glu Ile Met Leu Pro Val Phe Asp
 530 535 540

Ala Pro Gln Asn Leu Val Glu Gln Ala Lys Leu Thr Ala Ala Thr Asn
 545 550 555 560

Ala Lys Gln

<210> 22

<211> 33

<212> DNA

<213> artificial sequence

<220>

<223> artificial sequence

<400> 22

tctttaatta atgggttgct cgctattcat ata

33

<210> 23

<211> 32

<212> DNA

<213> . artificial sequence

<220>

<223> artificial sequence

<400> 23

ctaaagcttt taggccagag tggctcttgcg cg

32

<210> 24

<211> 1674

<212> DNA

<213> Acetobacter pasteurianus

<220>

<221> CDS

<222> (1)..(1674)

<223>

<400> 24

gtg acc tat act gtt ggc atg tat ctt gca gaa cgc ctt gta cag atc 48
Val Thr Tyr Thr Val Gly Met Tyr Leu Ala Glu Arg Leu Val Gln Ile
1 5 10 15

ggg ctg aag cat cac ttc gcc gtg ggc ggc gac tac aat ctc gtt ctt 96
Gly Leu Lys His His Phe Ala Val Gly Gly Asp Tyr Asn Leu Val Leu
20 25 30

ctg gat cag ttg ctc ctc aac aag gac atg aaa cag atc tat tgc tgc 144
Leu Asp Gln Leu Leu Leu Asn Lys Asp Met Lys Gln Ile Tyr Cys Cys
35 40 45

aat gag ttg aac tgt ggc ttc agc gcg gaa ggc tac gcc cgt tct aac 192
Asn Glu Leu Asn Cys Gly Phe Ser Ala Glu Gly Tyr Ala Arg Ser Asn
50 55 60

ggg gct gcg gca gcg gtt gtc acc ttc agc gtt ggc gcc att tcc gcc 240
Gly Ala Ala Ala Ala Val Val Thr Phe Ser Val Gly Ala Ile Ser Ala
65 70 75 80

atg aac gcc ctc ggc ggc gcc tat gcc gaa aac ctg ccg gtt atc ctg 288
Met Asn Ala Leu Gly Gly Ala Tyr Ala Glu Asn Leu Pro Val Ile Leu
85 90 95

att tcc gcc gcg ccc aac agc aat gat cag ggc aca ggt cat atc ctg 336
Ile Ser Gly Ala Pro Asn Ser Asn Asp Gln Gly Thr Gly His Ile Leu

100	105	110	
cat cac aca atc ggc aag acg gat tac agc tac cag ctt gaa atg gcc His His Thr Ile Gly Lys Thr Asp Tyr Ser Tyr Gln Leu Glu Met Ala 115 120 125			384
cgt cag gtc acc tgt gcc gcc gaa agc att acc gac gct cac tcc gcc Arg Gln Val Thr Cys Ala Ala Glu Ser Ile Thr Asp Ala His Ser Ala 130 135 140			432
ccg gcc aag att gac cac gtc att cgc acg gcg ctg cgc gag cgt aag Pro Ala Lys Ile Asp His Val Ile Arg Thr Ala Leu Arg Glu Arg Lys 145 150 155 160			480
ccg gcc tat ctg gac atc gcg tgc aac att gcc tcc gag ccc tgc gtg Pro Ala Tyr Leu Asp Ile Ala Cys Asn Ile Ala Ser Glu Pro Cys Val 165 170 175			528
cgg cct ggc cct gtc agc agc ctg ctg tcc gag cct gaa atc gac cac Arg Pro Gly Pro Val Ser Ser Leu Leu Ser Glu Pro Glu Ile Asp His 180 185 190			576
acg agc ctg aag gcc gca gtg gac gcc acg gtt gcc ttg ctg aaa aat Thr Ser Leu Lys Ala Ala Val Asp Ala Thr Val Ala Leu Leu Lys Asn 195 200 205			624
cgg cca gcc ccc gtc atg ctg ctg ggc agc aag ctg cgg gcc gcc aac Arg Pro Ala Pro Val Met Leu Leu Gly Ser Lys Leu Arg Ala Ala Asn 210 215 220			672
gca ctg gcc gca acc gaa acg ctg gca gac aag ctg caa tgc gcg gtg Ala Leu Ala Ala Thr Glu Thr Leu Ala Asp Lys Leu Gln Cys Ala Val 225 230 235 240			720
acc atc atg gcg gcc gcg aaa ggc ttt ttc ccc gaa gac cac gcg ggt Thr Ile Met Ala Ala Ala Lys Gly Phe Phe Pro Glu Asp His Ala Gly 245 250 255			768
ttc cgc ggc ctg tac tgg ggc gaa gtc tcg aac ccc ggc gtg cag gaa Phe Arg Gly Leu Tyr Trp Gly Glu Val Ser Asn Pro Gly Val Gln Glu 260 265 270			816
ctg gtg gag acc tcc gac gca ctg ctg tgc atc gcc ccc gta ttc aac Leu Val Glu Thr Ser Asp Ala Leu Leu Cys Ile Ala Pro Val Phe Asn 275 280 285			864
gac tat tca aca gtc ggc tgg tcg ggc atg ccc aag ggc ccc aat gtg Asp Tyr Ser Thr Val Gly Trp Ser Gly Met Pro Lys Gly Pro Asn Val 290 295 300			912
att ctg gct gag ccc gac cgc gta acg gtc gat ggc cgc gcc tat gac Ile Leu Ala Glu Pro Asp Arg Val Thr Val Asp Gly Arg Ala Tyr Asp 305 310 315 320			960
ggc ttt acc ctg cgc gcc ttc ctg cag gct ctg gcg gaa aaa gcc ccc Gly Phe Thr Leu Arg Ala Phe Leu Gln Ala Leu Ala Glu Lys Ala Pro 325 330 335			1008
gcg cgc ccg gcc tcc gca cag aaa agc agc gtc ccg acg tgc tcg ctc			1056

Ala Arg Pro Ala Ser Ala Gln Lys Ser Ser Val Pro Thr Cys Ser Leu	
340 345 350	
acc gcg aca tcc gat gaa gcc ggt ctg acg aat gac gaa atc gtc cgt	1104
Thr Ala Thr Ser Asp Glu Ala Gly Leu Thr Asn Asp Glu Ile Val Arg	
355 360 365	
cat atc aac gcc ctg ctg aca tca aac acg acg ctg gtg gca gaa acc	1152
His Ile Asn Ala Leu Leu Thr Ser Asn Thr Thr Leu Val Ala Glu Thr	
370 375 380	
ggc gat tca tgg ttc aat gcc atg cgc atg acc ctg gcc ggt gcg cgc	1200
Gly Asp Ser Trp Phe Asn Ala Met Arg Met Thr Leu Ala Gly Ala Arg	
385 390 395 400	
gtg gaa ctg gaa atg cag tgg ggc cat atc ggc tgg tcc gtg ccc tcc	1248
Val Glu Leu Glu Met Gln Trp Gly His Ile Gly Trp Ser Val Pro Ser	
405 410 415	
gcg ttc ggc aat gcc atg ggc tgc cag gac cgc cag cat gtg gtg atg	1296
Ala Phe Gly Asn Ala Met Gly Ser Gln Asp Arg Gln His Val Val Met	
420 425 430	
gta ggc gat ggc tcc ttc cag ctt acc gcg cag gaa gtg gct cag atg	1344
Val Gly Asp Gly Ser Phe Gln Leu Thr Ala Gln Glu Val Ala Gln Met	
435 440 445	
gtg cgc tac gaa ctg ccc gtc att atc ttt ctg atc aac aac cgt ggc	1392
Val Arg Tyr Glu Leu Pro Val Ile Ile Phe Leu Ile Asn Asn Arg Gly	
450 455 460	
tat gtc att gaa atc gcc att cat gac ggc ccg tac aac tat atc aag	1440
Tyr Val Ile Glu Ile Ala Ile His Asp Gly Pro Tyr Asn Tyr Ile Lys	
465 470 475 480	
aac tgg gat tac gcc ggc ctg atg gaa gtc ttc aac gcc gga gaa ggc	1488
Asn Trp Asp Tyr Ala Gly Leu Met Glu Val Phe Asn Ala Gly Glu Gly	
485 490 495	
cat gga ctt ggc ctg aaa gcc acc acc ccg aag gaa ctg aca gaa gcc	1536
His Gly Leu Gly Leu Lys Ala Thr Thr Pro Lys Glu Leu Thr Glu Ala	
500 505 510	
atc gcc agg gca aaa gcc aat acc cgc ggc ccg acg ctg atc gaa tgc	1584
Ile Ala Arg Ala Lys Ala Asn Thr Arg Gly Pro Thr Leu Ile Glu Cys	
515 520 525	
cag atc gac cgc acg gac tgc acg gat atg ctg gtt caa tgg ggc cgc	1632
Gln Ile Asp Arg Thr Asp Cys Thr Asp Met Leu Val Gln Trp Gly Arg	
530 535 540	
aag gtt gcc tca acc aac gcg cgc aag acc act ctg gcc tga	1674
Lys Val Ala Ser Thr Asn Ala Arg Lys Thr Thr Leu Ala	
545 550 555	

<210> 25

<211> 557

<212> PRT

<213> Acetobacter pasteurianus

<400> 25

Val Thr Tyr Thr Val Gly Met Tyr Leu Ala Glu Arg Leu Val Gln Ile
 1 5 10 15

Gly Leu Lys His His Phe Ala Val Gly Gly Asp Tyr Asn Leu Val Leu
 20 25 30

Leu Asp Gln Leu Leu Leu Asn Lys Asp Met Lys Gln Ile Tyr Cys Cys
 35 40 45

Asn Glu Leu Asn Cys Gly Phe Ser Ala Glu Gly Tyr Ala Arg Ser Asn
 50 55 60

Gly Ala Ala Ala Ala Val Val Thr Phe Ser Val Gly Ala Ile Ser Ala
 65 70 75 80

Met Asn Ala Leu Gly Gly Ala Tyr Ala Glu Asn Leu Pro Val Ile Leu
 85 90 95

Ile Ser Gly Ala Pro Asn Ser Asn Asp Gln Gly Thr Gly His Ile Leu
 100 105 110

His His Thr Ile Gly Lys Thr Asp Tyr Ser Tyr Gln Leu Glu Met Ala
 115 120 125

Arg Gln Val Thr Cys Ala Ala Glu Ser Ile Thr Asp Ala His Ser Ala
 130 135 140

Pro Ala Lys Ile Asp His Val Ile Arg Thr Ala Leu Arg Glu Arg Lys
 145 150 155 160

Pro Ala Tyr Leu Asp Ile Ala Cys Asn Ile Ala Ser Glu Pro Cys Val
 165 170 175

Arg Pro Gly Pro Val Ser Ser Leu Leu Ser Glu Pro Glu Ile Asp His
 180 185 190

Thr Ser Leu Lys Ala Ala Val Asp Ala Thr Val Ala Leu Leu Lys Asn
 195 200 205

Arg Pro Ala Pro Val Met Leu Leu Gly Ser Lys Leu Arg Ala Ala Asn
 210 215 220

Ala Leu Ala Ala Thr Glu Thr Leu Ala Asp Lys Leu Gln Cys Ala Val
 225 230 235 240

Thr Ile Met Ala Ala Ala Lys Gly Phe Phe Pro Glu Asp His Ala Gly
 245 250 255

Phe Arg Gly Leu Tyr Trp Gly Glu Val Ser Asn Pro Gly Val Gln Glu
 260 265 270

Leu Val Glu Thr Ser Asp Ala Leu Leu Cys Ile Ala Pro Val Phe Asn
 275 280 285

Asp Tyr Ser Thr Val Gly Trp Ser Gly Met Pro Lys Gly Pro Asn Val
 290 295 300

Ile Leu Ala Glu Pro Asp Arg Val Thr Val Asp Gly Arg Ala Tyr Asp
 305 310 315 320

Gly Phe Thr Leu Arg Ala Phe Leu Gln Ala Leu Ala Glu Lys Ala Pro
 325 330 335

Ala Arg Pro Ala Ser Ala Gln Lys Ser Ser Val Pro Thr Cys Ser Leu
 340 345 350

Thr Ala Thr Ser Asp Glu Ala Gly Leu Thr Asn Asp Glu Ile Val Arg
 355 360 365

His Ile Asn Ala Leu Leu Thr Ser Asn Thr Thr Leu Val Ala Glu Thr
 370 375 380

Gly Asp Ser Trp Phe Asn Ala Met Arg Met Thr Leu Ala Gly Ala Arg
 385 390 395 400

Val Glu Leu Glu Met Gln Trp Gly His Ile Gly Trp Ser Val Pro Ser
 405 410 415

Ala Phe Gly Asn Ala Met Gly Ser Gln Asp Arg Gln His Val Val Met
 420 425 430

Val Gly Asp Gly Ser Phe Gln Leu Thr Ala Gln Glu Val Ala Gln Met

435

440

445

Val Arg Tyr Glu Leu Pro Val Ile Ile Phe Leu Ile Asn Asn Arg Gly
 450 455 460

Tyr Val Ile Glu Ile Ala Ile His Asp Gly Pro Tyr Asn Tyr Ile Lys
 465 470 475 480

Asn Trp Asp Tyr Ala Gly Leu Met Glu Val Phe Asn Ala Gly Glu Gly
 485 490 495

His Gly Leu Gly Leu Lys Ala Thr Thr Pro Lys Glu Leu Thr Glu Ala
 500 505 510

Ile Ala Arg Ala Lys Ala Asn Thr Arg Gly Pro Thr Leu Ile Glu Cys
 515 520 525

Gln Ile Asp Arg Thr Asp Cys Thr Asp Met Leu Val Gln Trp Gly Arg
 530 535 540

Lys Val Ala Ser Thr Asn Ala Arg Lys Thr Thr Leu Ala
 545 550 555

<210> 26

<211> 32

<212> DNA

<213> artificial sequence

<220>

<223> artificial sequence

<400> 26

atcttaatta atgtataccg ttggtatgta ct

32

<210> 27

<211> 34

<212> DNA

<213> artificial sequence

<220>

<223> artificial sequence

<400> 27

tatgcgggccc cttacgcttg tggtttgcca gagt

34

<210> 28

<211> 1671

<212> DNA

<213> Zymobacter palmae

<220>

<221> CDS

<222> (1)..(1671)

<223>

<400> 28

atg	tat	acc	gtt	ggt	atg	tac	ttg	gca	gaa	cgc	cta	gcc	cag	atc	ggc	48
Met	Tyr	Thr	Val	Gly	Met	Tyr	Leu	Ala	Glu	Arg	Leu	Ala	Gln	Ile	Gly	
1			5					10					15			

ctg	aaa	cac	cac	ttt	gcc	gtg	gcc	ggt	gac	tac	aac	ctg	gtg	ttg	ctt	96
Leu	Lys	His	His	Phe	Ala	Val	Ala	Gly	Asp	Tyr	Asn	Leu	Val	Leu	Leu	
		20					25					30				

gat	cag	ctc	ctg	ctg	aac	aaa	gac	atg	gag	cag	gtc	tac	tgc	tgt	aac	144
Asp	Gln	Leu	Leu	Leu	Asn	Lys	Asp	Met	Glu	Gln	Val	Tyr	Cys	Cys	Asn	
	35					40						45				

gaa	ctt	aac	tgc	ggc	ttt	agc	gcc	gaa	ggt	tac	gct	cgt	gca	cgt	ggt	192
Glu	Leu	Asn	Cys	Gly	Phe	Ser	Ala	Glu	Gly	Tyr	Ala	Arg	Ala	Arg	Gly	
	50					55				60						

gcc	gcc	gct	gcc	atc	gtc	acg	ttc	agc	gta	ggt	gct	atc	tct	gca	atg	240
Ala	Ala	Ala	Ala	Ile	Val	Thr	Phe	Ser	Val	Gly	Ala	Ile	Ser	Ala	Met	
65				70					75					80		

aac	gcc	atc	ggt	ggc	gcc	tat	gca	gaa	aac	ctg	ccg	gtc	atc	ctg	atc	288
Asn	Ala	Ile	Gly	Gly	Ala	Tyr	Ala	Glu	Asn	Leu	Pro	Val	Ile	Leu	Ile	
			85						90					95		

tct	ggc	tca	ccg	aac	acc	aat	gac	tac	ggc	aca	ggc	cac	atc	ctg	cac	336
Ser	Gly	Ser	Pro	Asn	Thr	Asn	Asp	Tyr	Gly	Thr	Gly	His	Ile	Leu	His	
			100					105					110			

cac	acc	att	ggt	act	act	gac	tat	aac	tat	cag	ctg	gaa	atg	gta	aaa	384
His	Thr	Ile	Gly	Thr	Thr	Asp	Tyr	Asn	Tyr	Gln	Leu	Glu	Met	Val	Lys	

115	120	125	
cac gtt acc tgc gca cgt gaa agc atc gtt tct gcc gaa gaa gca ccg His Val Thr Cys Ala Arg Glu Ser Ile Val Ser Ala Glu Glu Ala Pro 130 135 140			432
gca aaa atc gac cac gtc atc cgt acg gct cta cgt gaa cgc aaa ccg Ala Lys Ile Asp His Val Ile Arg Thr Ala Leu Arg Glu Arg Lys Pro 145 150 155 160			480
gct tat ctg gaa atc gca tgc aac gtc gct gcc gct gaa tgt gtt cgt Ala Tyr Leu Glu Ile Ala Cys Asn Val Ala Gly Ala Glu Cys Val Arg 165 170 175			528
ccg gcc ccg atc aat agc ctg ctg cgt gaa ctc gaa gtt gac cag acc Pro Gly Pro Ile Asn Ser Leu Leu Arg Glu Leu Glu Val Asp Gln Thr 180 185 190			576
agt gtc act gcc gct gta gat gcc gcc gta gaa tgg ctg cag gac cgc Ser Val Thr Ala Ala Val Asp Ala Ala Val Glu Trp Leu Gln Asp Arg 195 200 205			624
cag aac gtc gtc atg ctg gtc ggt agc aaa ctg cgt gcc gct gcc gct Gln Asn Val Val Met Leu Val Gly Ser Lys Leu Arg Ala Ala Ala Ala 210 215 220			672
gaa aaa cag gct gtt gcc cta gcg gac cgc ctg gcc tgc gct gtc acg Glu Lys Gln Ala Val Ala Leu Ala Asp Arg Leu Gly Cys Ala Val Thr 225 230 235 240			720
atc atg gct gcc gaa aaa gcc ttc ttc ccg gaa gat cat ccg aac ttc Ile Met Ala Ala Glu Lys Gly Phe Phe Pro Glu Asp His Pro Asn Phe 245 250 255			768
cgc gcc ctg tac tgg ggt gaa gtc agc tcc gaa ggt gca cag gaa ctg Arg Gly Leu Tyr Trp Gly Glu Val Ser Ser Glu Gly Ala Gln Glu Leu 260 265 270			816
gtt gaa aac gcc gat gcc atc ctg tgt ctg gca ccg gta ttc aac gac Val Glu Asn Ala Asp Ala Ile Leu Cys Leu Ala Pro Val Phe Asn Asp 275 280 285			864
tat gct acc gtt gcc tgg aac tcc tgg ccg aaa gcc gac aat gtc atg Tyr Ala Thr Val Gly Trp Asn Ser Trp Pro Lys Gly Asp Asn Val Met 290 295 300			912
gtc atg gac acc gac cgc gtc act ttc gca gga cag tcc ttc gaa ggt Val Met Asp Thr Asp Arg Val Thr Phe Ala Gly Gln Ser Phe Glu Gly 305 310 315 320			960
ctg tca ttg agc acc ttc gcc gca gca ctg gct gag aaa gca cct tct Leu Ser Leu Ser Thr Phe Ala Ala Ala Leu Ala Glu Lys Ala Pro Ser 325 330 335			1008
cgc ccg gca acg act caa gcc act caa gca ccg gta ctg ggt att gag Arg Pro Ala Thr Thr Gln Gly Thr Gln Ala Pro Val Leu Gly Ile Glu 340 345 350			1056
gcc gca gag ccc aat gca ccg ctg acc aat gac gaa atg acg cgt cag			1104

Ala Ala Glu Pro Asn Ala Pro Leu Thr Asn Asp Glu Met Thr Arg Gln
 355 360 365

atc cag tgc ctg atc act tcc gac act act ctg aca gca gaa aca ggt 1152
 Ile Gln Ser Leu Ile Thr Ser Asp Thr Thr Leu Thr Ala Glu Thr Gly
 370 375 380

gac tct tgg ttc aac gct tct cgc atg ccg att cct ggc ggt gct cgt 1200
 Asp Ser Trp Phe Asn Ala Ser Arg Met Pro Ile Pro Gly Gly Ala Arg
 385 390 395 400

gtc gaa ctg gaa atg caa tgg ggt cat atc ggt tgg tcc gta cct tct 1248
 Val Glu Leu Glu Met Gln Trp Gly His Ile Gly Trp Ser Val Pro Ser
 405 410 415

gca ttc ggt aac gcc gtt ggt tct ccg gag cgt cgc cac atc atg atg 1296
 Ala Phe Gly Asn Ala Val Gly Ser Pro Glu Arg Arg His Ile Met Met
 420 425 430

gtc ggt gat ggc tct ttc cag ctg act gct caa gaa gtt gct cag atg 1344
 Val Gly Asp Gly Ser Phe Gln Leu Thr Ala Gln Glu Val Ala Gln Met
 435 440 445

atc cgc tat gaa atc ccg gtc atc atc ttc ctg atc aac aac cgc ggt 1392
 Ile Arg Tyr Glu Ile Pro Val Ile Ile Phe Leu Ile Asn Asn Arg Gly
 450 455 460

tac gtc atc gaa atc gct atc cat gac ggc cct tac aac tac atc aaa 1440
 Tyr Val Ile Glu Ile Ala Ile His Asp Gly Pro Tyr Asn Tyr Ile Lys
 465 470 475 480

aac tgg aac tac gct ggc ctg atc gac gtc ttc aat gac gaa gat ggt 1488
 Asn Trp Asn Tyr Ala Gly Leu Ile Asp Val Phe Asn Asp Glu Asp Gly
 485 490 495

cat ggc ctg ggt ctg aaa gct tct act ggt gca gaa cta gaa ggc gct 1536
 His Gly Leu Gly Leu Lys Ala Ser Thr Gly Ala Glu Leu Glu Gly Ala
 500 505 510

atc aag aaa gca ctc gac aat cgt cgc ggt ccg acg ctg atc gaa tgt 1584
 Ile Lys Lys Ala Leu Asp Asn Arg Arg Gly Pro Thr Leu Ile Glu Cys
 515 520 525

aac atc gct cag gac gac tgc act gaa acc ctg att gct tgg ggt aaa 1632
 Asn Ile Ala Gln Asp Asp Cys Thr Glu Thr Leu Ile Ala Trp Gly Lys
 530 535 540

cgt gta gca gct acc aac tct cgc aaa cca caa gcg taa 1671
 Arg Val Ala Ala Thr Asn Ser Arg Lys Pro Gln Ala
 545 550 555

<210> 29

<211> 556

<212> PRT

<213> Zymobacter palmae

<400> 29

Met Tyr Thr Val Gly Met Tyr Leu Ala Glu Arg Leu Ala Gln Ile Gly
 1 5 10 15

Leu Lys His His Phe Ala Val Ala Gly Asp Tyr Asn Leu Val Leu Leu
 20 25 30

Asp Gln Leu Leu Leu Asn Lys Asp Met Glu Gln Val Tyr Cys Cys Asn
 35 40 45

Glu Leu Asn Cys Gly Phe Ser Ala Glu Gly Tyr Ala Arg Ala Arg Gly
 50 55 60

Ala Ala Ala Ala Ile Val Thr Phe Ser Val Gly Ala Ile Ser Ala Met
 65 70 75 80

Asn Ala Ile Gly Gly Ala Tyr Ala Glu Asn Leu Pro Val Ile Leu Ile
 85 90 95

Ser Gly Ser Pro Asn Thr Asn Asp Tyr Gly Thr Gly His Ile Leu His
 100 105 110

His Thr Ile Gly Thr Thr Asp Tyr Asn Tyr Gln Leu Glu Met Val Lys
 115 120 125

His Val Thr Cys Ala Arg Glu Ser Ile Val Ser Ala Glu Glu Ala Pro
 130 135 140

Ala Lys Ile Asp His Val Ile Arg Thr Ala Leu Arg Glu Arg Lys Pro
 145 150 155 160

Ala Tyr Leu Glu Ile Ala Cys Asn Val Ala Gly Ala Glu Cys Val Arg
 165 170 175

Pro Gly Pro Ile Asn Ser Leu Leu Arg Glu Leu Glu Val Asp Gln Thr
 180 185 190

Ser Val Thr Ala Ala Val Asp Ala Ala Val Glu Trp Leu Gln Asp Arg
 195 200 205

Gln Asn Val Val Met Leu Val Gly Ser Lys Leu Arg Ala Ala Ala Ala
 210 215 220

Glu Lys Gln Ala Val Ala Leu Ala Asp Arg Leu Gly Cys Ala Val Thr
 225 230 235 240

Ile Met Ala Ala Glu Lys Gly Phe Phe Pro Glu Asp His Pro Asn Phe
 245 250 255

Arg Gly Leu Tyr Trp Gly Glu Val Ser Ser Glu Gly Ala Gln Glu Leu
 260 265 270

Val Glu Asn Ala Asp Ala Ile Leu Cys Leu Ala Pro Val Phe Asn Asp
 275 280 285

Tyr Ala Thr Val Gly Trp Asn Ser Trp Pro Lys Gly Asp Asn Val Met
 290 295 300

Val Met Asp Thr Asp Arg Val Thr Phe Ala Gly Gln Ser Phe Glu Gly
 305 310 315 320

Leu Ser Leu Ser Thr Phe Ala Ala Ala Leu Ala Glu Lys Ala Pro Ser
 325 330 335

Arg Pro Ala Thr Thr Gln Gly Thr Gln Ala Pro Val Leu Gly Ile Glu
 340 345 350

Ala Ala Glu Pro Asn Ala Pro Leu Thr Asn Asp Glu Met Thr Arg Gln
 355 360 365

Ile Gln Ser Leu Ile Thr Ser Asp Thr Thr Leu Thr Ala Glu Thr Gly
 370 375 380

Asp Ser Trp Phe Asn Ala Ser Arg Met Pro Ile Pro Gly Gly Ala Arg
 385 390 395 400

Val Glu Leu Glu Met Gln Trp Gly His Ile Gly Trp Ser Val Pro Ser
 405 410 415

Ala Phe Gly Asn Ala Val Gly Ser Pro Glu Arg Arg His Ile Met Met
 420 425 430

Val Gly Asp Gly Ser Phe Gln Leu Thr Ala Gln Glu Val Ala Gln Met
 435 440 445

Ile Arg Tyr Glu Ile Pro Val Ile Ile Phe Leu Ile Asn Asn Arg Gly

450 455 460

Tyr Val Ile Glu Ile Ala Ile His Asp Gly Pro Tyr Asn Tyr Ile Lys
465 470 475 480

Asn Trp Asn Tyr Ala Gly Leu Ile Asp Val Phe Asn Asp Glu Asp Gly
485 490 495

His Gly Leu Gly Leu Lys Ala Ser Thr Gly Ala Glu Leu Glu Gly Ala
500 505 510

Ile Lys Lys Ala Leu Asp Asn Arg Arg Gly Pro Thr Leu Ile Glu Cys
515 520 525

Asn Ile Ala Gln Asp Asp Cys Thr Glu Thr Leu Ile Ala Trp Gly Lys
530 535 540

Arg Val Ala Ala Thr Asn Ser Arg Lys Pro Gln Ala
545 550 555

<210> 30

<211> 32

<212> DNA

<213> artificial sequence

<400> 30

ctattaatta atggcttcgg tacacggcac ca

32

<210> 31

<211> 34

<212> DNA

<213> artificial sequence

<400> 31

tatgcggccg cttacttcac cgggcttacg gtgc

34

<210> 32

<211> 1587

<212> DNA

<213> Pseudomonas putida

<220>

<221> CDS

<222> (1)..(1584)

<223>

<400> 32

atg gct tcg gta cac ggc acc aca tac gaa ctc ttg cga cgt caa ggc	48
Met Ala Ser Val His Gly Thr Thr Tyr Glu Leu Leu Arg Arg Gln Gly	
1 5 10 15	

atc gat acg gtc ttc ggc aat cct ggc tcg aac gag ctc ccg ttt ttg	96
Ile Asp Thr Val Phe Gly Asn Pro Gly Ser Asn Glu Leu Pro Phe Leu	
20 25 30	

aag gac ttt cca gag gac ttt cga tac atc ctg gct ttg cag gaa gcg	144
Lys Asp Phe Pro Glu Asp Phe Arg Tyr Ile Leu Ala Leu Gln Glu Ala	
35 40 45	

tgt gtg gtg ggc att gca gac ggc tat gcg caa gcc agt cgg aag ccg	192
Cys Val Val Gly Ile Ala Asp Gly Tyr Ala Gln Ala Ser Arg Lys Pro	
50 55 60	

gct ttc att aac ctg cat tct gct gct ggt acc ggc aat gct atg ggt	240
Ala Phe Ile Asn Leu His Ser Ala Ala Gly Thr Gly Asn Ala Met Gly	
65 70 75 80	

gca ctc agt aac gcc tgg aac tca cat tcc ccg ctg atc gtc act gcc	288
Ala Leu Ser Asn Ala Trp Asn Ser His Ser Pro Leu Ile Val Thr Ala	
85 90 95	

ggc cag cag acc agg gcg atg att ggc gtt gaa gct ctg ctg acc aac	336
Gly Gln Gln Thr Arg Ala Met Ile Gly Val Glu Ala Leu Leu Thr Asn	
100 105 110	

gtc gat gcc gcc aac ctg cca cga cca ctt gtc aaa tgg agc tac gag	384
Val Asp Ala Ala Asn Leu Pro Arg Pro Leu Val Lys Trp Ser Tyr Glu	
115 120 125	

ccc gca agc gca gca gaa gtc cct cat gcg atg agc agg gct atc cat	432
Pro Ala Ser Ala Ala Glu Val Pro His Ala Met Ser Arg Ala Ile His	
130 135 140	

atg gca agc atg gcg cca caa ggc cct gtc tat ctt tcg gtg cca tat	480
Met Ala Ser Met Ala Pro Gln Gly Pro Val Tyr Leu Ser Val Pro Tyr	
145 150 155 160	

gac gat tgg gat aag gat gct gat cct cag tcc cac cac ctt ttt gat	528
Asp Asp Trp Asp Lys Asp Ala Asp Pro Gln Ser His His Leu Phe Asp	

	165	170	175	
cgc cat gtc agt tca tca gta cgc ctg aac gac cag gat ctc gat att				576
Arg His Val Ser Ser Ser Val Arg Leu Asn Asp Gln Asp Leu Asp Ile				
	180	185	190	
ctg gtg aaa gct ctc aac agc gca tcc aac ccg gcg atc gtc ctg ggc				624
Leu Val Lys Ala Leu Asn Ser Ala Ser Asn Pro Ala Ile Val Leu Gly				
	195	200	205	
ccg gac gtc gac gca gca aat gcg aac gca gac tgc gtc atg ttg gcc				672
Pro Asp Val Asp Ala Ala Asn Ala Asn Ala Asp Cys Val Met Leu Ala				
	210	215	220	
gaa cgc ctc aaa gct ccg gtt tgg gtt gcg cca tcc gct cca cgc tgc				720
Glu Arg Leu Lys Ala Pro Val Trp Val Ala Pro Ser Ala Pro Arg Cys				
	225	230	235	240
cca ttc cct acc cgt cat cct tgc ttc cgt gga ttg atg cca gct ggc				768
Pro Phe Pro Thr Arg His Pro Cys Phe Arg Gly Leu Met Pro Ala Gly				
	245	250	255	
atc gca gcg att tct cag ctg ctc gaa ggt cac gat gtg gtt ttg gta				816
Ile Ala Ala Ile Ser Gln Leu Leu Glu Gly His Asp Val Val Leu Val				
	260	265	270	
atc ggc gct cca gtg ttc cgt tac cac caa tac gac cca ggt caa tat				864
Ile Gly Ala Pro Val Phe Arg Tyr His Gln Tyr Asp Pro Gly Gln Tyr				
	275	280	285	
ctc aaa cct ggc acg cga ttg att tcg gtg acc tgc gac ccg ctc gaa				912
Leu Lys Pro Gly Thr Arg Leu Ile Ser Val Thr Cys Asp Pro Leu Glu				
	290	295	300	
gct gca cgc gcg cca atg ggc gat gcg atc gtg gca gac att ggt gcg				960
Ala Ala Arg Ala Pro Met Gly Asp Ala Ile Val Ala Asp Ile Gly Ala				
	305	310	315	320
atg gct agc gct ctt gcc aac ttg gtt gaa gag agc agc cgc cag ctc				1008
Met Ala Ser Ala Leu Ala Asn Leu Val Glu Glu Ser Ser Arg Gln Leu				
	325	330	335	
cca act gca gct ccg gaa ccc gcg aag gtt gac caa gac gct ggc cga				1056
Pro Thr Ala Ala Pro Glu Pro Ala Lys Val Asp Gln Asp Ala Gly Arg				
	340	345	350	
ctt cac cca gag aca gtg ttc gac aca ctg aac gac atg gcc ccg gag				1104
Leu His Pro Glu Thr Val Phe Asp Thr Leu Asn Asp Met Ala Pro Glu				
	355	360	365	
aat gcg att tac ctg aac gag tcg act tca acg acc gcc caa atg tgg				1152
Asn Ala Ile Tyr Leu Asn Glu Ser Thr Ser Thr Thr Ala Gln Met Trp				
	370	375	380	
cag cgc ctg aac atg cgc aac cct ggt agc tac tac ttc tgt gca gct				1200
Gln Arg Leu Asn Met Arg Asn Pro Gly Ser Tyr Tyr Phe Cys Ala Ala				
	385	390	395	400
ggc gga ctg ggc ttc gcc ctg cct gca gca att ggc gtt caa ctc gca				1248

Gly Gly Leu Gly Phe Ala Leu Pro Ala Ala Ile Gly Val Gln Leu Ala
 405 410 415

gaa ccc gag cga caa gtc atc gcc gtc att ggc gac gga tcg gcg aac 1296
 Glu Pro Glu Arg Gln Val Ile Ala Val Ile Gly Asp Gly Ser Ala Asn
 420 425 430

tac agc att agt gcg ttg tgg act gca gct cag tac aac atc ccc act 1344
 Tyr Ser Ile Ser Ala Leu Trp Thr Ala Ala Gln Tyr Asn Ile Pro Thr
 435 440 445

atc ttc gtg atc atg aac aac ggc acc tac ggt gcg ttg cga tgg ttt 1392
 Ile Phe Val Ile Met Asn Asn Gly Thr Tyr Gly Ala Leu Arg Trp Phe
 450 455 460

gcc ggc gtt ctc gaa gca gaa aac gtt cct ggg ctg gat gtg cca ggg 1440
 Ala Gly Val Leu Glu Ala Glu Asn Val Pro Gly Leu Asp Val Pro Gly
 465 470 475 480

atc gac ttc cgc gca ctc gcc aag ggc tat ggt gtc caa gcg ctg aaa 1488
 Ile Asp Phe Arg Ala Leu Ala Lys Gly Tyr Gly Val Gln Ala Leu Lys
 485 490 495

gcc gac aac ctt gag cag ctc aag ggt tcg cta caa gaa gcg ctt tct 1536
 Ala Asp Asn Leu Glu Gln Leu Lys Gly Ser Leu Gln Glu Ala Leu Ser
 500 505 510

gcc aaa ggc ccg gta ctt atc gaa gta agc acc gta agc ccg gtg aag 1584
 Ala Lys Gly Pro Val Leu Ile Glu Val Ser Thr Val Ser Pro Val Lys
 515 520 525

tga 1587

<210> 33

<211> 528

<212> PRT

<213> *Pseudomonas putida*

<400> 33

Met Ala Ser Val His Gly Thr Thr Tyr Glu Leu Leu Arg Arg Gln Gly
 1 5 10 15

Ile Asp Thr Val Phe Gly Asn Pro Gly Ser Asn Glu Leu Pro Phe Leu
 20 25 30

Lys Asp Phe Pro Glu Asp Phe Arg Tyr Ile Leu Ala Leu Gln Glu Ala
 35 40 45

Cys Val Val Gly Ile Ala Asp Gly Tyr Ala Gln Ala Ser Arg Lys Pro

50 55 60
 Ala Phe Ile Asn Leu His Ser Ala Ala Gly Thr Gly Asn Ala Met Gly
 65 70 75 80
 Ala Leu Ser Asn Ala Trp Asn Ser His Ser Pro Leu Ile Val Thr Ala
 85 90 95
 Gly Gln Gln Thr Arg Ala Met Ile Gly Val Glu Ala Leu Leu Thr Asn
 100 105 110
 Val Asp Ala Ala Asn Leu Pro Arg Pro Leu Val Lys Trp Ser Tyr Glu
 115 120 125
 Pro Ala Ser Ala Ala Glu Val Pro His Ala Met Ser Arg Ala Ile His
 130 135 140
 Met Ala Ser Met Ala Pro Gln Gly Pro Val Tyr Leu Ser Val Pro Tyr
 145 150 155 160
 Asp Asp Trp Asp Lys Asp Ala Asp Pro Gln Ser His His Leu Phe Asp
 165 170 175
 Arg His Val Ser Ser Ser Val Arg Leu Asn Asp Gln Asp Leu Asp Ile
 180 185 190
 Leu Val Lys Ala Leu Asn Ser Ala Ser Asn Pro Ala Ile Val Leu Gly
 195 200 205
 Pro Asp Val Asp Ala Ala Asn Ala Asn Ala Asp Cys Val Met Leu Ala
 210 215 220
 Glu Arg Leu Lys Ala Pro Val Trp Val Ala Pro Ser Ala Pro Arg Cys
 225 230 235 240
 Pro Phe Pro Thr Arg His Pro Cys Phe Arg Gly Leu Met Pro Ala Gly
 245 250 255
 Ile Ala Ala Ile Ser Gln Leu Leu Glu Gly His Asp Val Val Leu Val
 260 265 270
 Ile Gly Ala Pro Val Phe Arg Tyr His Gln Tyr Asp Pro Gly Gln Tyr
 275 280 285

Leu Lys Pro Gly Thr Arg Leu Ile Ser Val Thr Cys Asp Pro Leu Glu
 290 295 300

Ala Ala Arg Ala Pro Met Gly Asp Ala Ile Val Ala Asp Ile Gly Ala
 305 310 315 320

Met Ala Ser Ala Leu Ala Asn Leu Val Glu Glu Ser Ser Arg Gln Leu
 325 330 335

Pro Thr Ala Ala Pro Glu Pro Ala Lys Val Asp Gln Asp Ala Gly Arg
 340 345 350

Leu His Pro Glu Thr Val Phe Asp Thr Leu Asn Asp Met Ala Pro Glu
 355 360 365

Asn Ala Ile Tyr Leu Asn Glu Ser Thr Ser Thr Thr Ala Gln Met Trp
 370 375 380

Gln Arg Leu Asn Met Arg Asn Pro Gly Ser Tyr Tyr Phe Cys Ala Ala
 385 390 395 400

Gly Gly Leu Gly Phe Ala Leu Pro Ala Ala Ile Gly Val Gln Leu Ala
 405 410 415

Glu Pro Glu Arg Gln Val Ile Ala Val Ile Gly Asp Gly Ser Ala Asn
 420 425 430

Tyr Ser Ile Ser Ala Leu Trp Thr Ala Ala Gln Tyr Asn Ile Pro Thr
 435 440 445

Ile Phe Val Ile Met Asn Asn Gly Thr Tyr Gly Ala Leu Arg Trp Phe
 450 455 460

Ala Gly Val Leu Glu Ala Glu Asn Val Pro Gly Leu Asp Val Pro Gly
 465 470 475 480

Ile Asp Phe Arg Ala Leu Ala Lys Gly Tyr Gly Val Gln Ala Leu Lys
 485 490 495

Ala Asp Asn Leu Glu Gln Leu Lys Gly Ser Leu Gln Glu Ala Leu Ser
 500 505 510

Ala Lys Gly Pro Val Leu Ile Glu Val Ser Thr Val Ser Pro Val Lys
 515 520 525

INTERNATIONAL SEARCH REPORT

International Application No.
PCT/EP2004/006848

A. CLASSIFICATION OF SUBJECT MATTER IPC 7 C07H7/027 C07H1/00 C12P7/42 C12N9/88 C12N1/20		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) IPC 7 C07H C12P C12N		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practical, search terms used) EPO-Internal, BEILSTEIN Data, WPI Data		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	SHELTON M C. ET AL: "2-Keto-3-deoxy-6-phosphogluconate aldolases as catalysts for stereocontrolled carbon-carbon bond formation" JOURNAL OF THE AMERICAN CHEMICAL SOCIETY, AMERICAN CHEMICAL SOCIETY, WASHINGTON, DC, US, vol. 118, no. 9, 6 March 1996 (1996-03-06), pages 2117-2125, XP002263455 ISSN: 0002-7863 Scheme 6	1
A	US 5 872 247 A (DUFLOT PIERRICK ET AL) 16 February 1999 (1999-02-16) examples	1
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<input checked="" type="checkbox"/> Further documents are listed in the continuation of box C. <input checked="" type="checkbox"/> Patent family members are listed in annex.		
* Special categories of cited documents:		
<div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> <p>*A* document defining the general state of the art which is not considered to be of particular relevance</p> <p>*E* earlier document but published on or after the international filing date</p> <p>*L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>*O* document referring to an oral disclosure, use, exhibition or other means</p> <p>*P* document published prior to the international filing date but later than the priority date claimed</p> </div> <div style="width: 45%;"> <p>*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</p> <p>*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>*&* document member of the same patent family</p> </div> </div>		
Date of the actual completion of the international search	Date of mailing of the international search report	
16 November 2004	29/11/2004	
Name and mailing address of the ISA	Authorized officer	
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax (+31-70) 340-3016	de Nooy, A	

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